

The Microsensor Company



Field Studies



February 2022
Tage Dalsgaard

Online Biogeochemistry Workshop



Wednesday 23 February

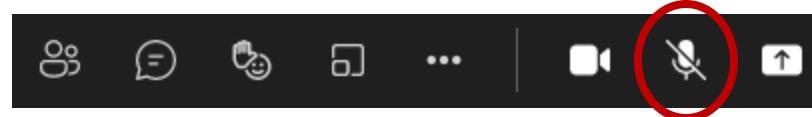
14:30-15:30 CET - Introduction to Microsensors
15:45-16:45 CET - Lab-based Studies

Thursday 24 February

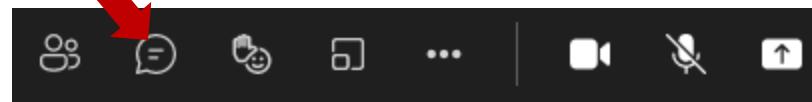
14:30-15:30 CET - Field Studies
15:45-16:30 CET - Demonstration of Field Microprofiling System
16:45-17:30 CET - Demonstration of Activity Calculation - Software

A few rules before we get started

1. Please turn off your microphone



2. Questions: During lecture please use chat.
After lecture you can unmute and ask.



Very application-specific questions may be better answered in a private session afterwards.

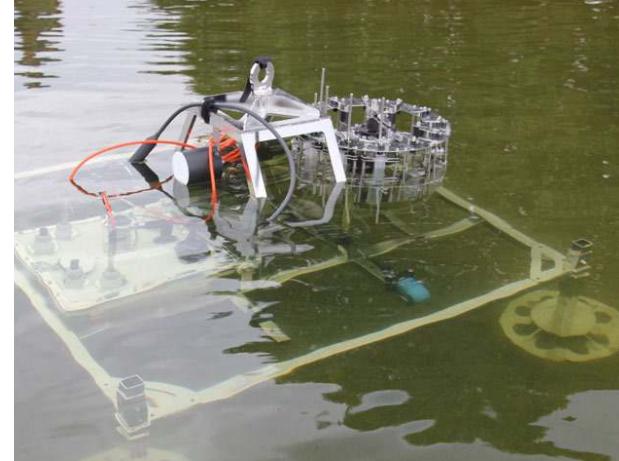
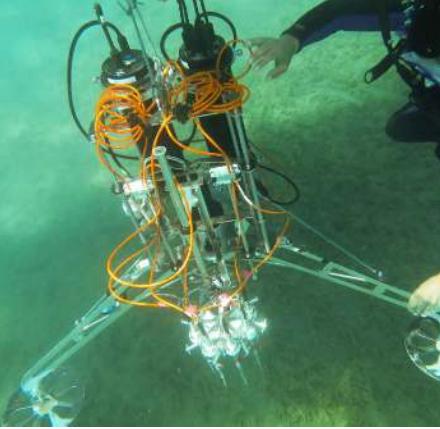
You will get access to all the presentations as PDF's + recordings shortly after the workshop.

Outline



- In situ UniAmp amplifiers and pressure compensation
- Microprofiles in situ - MiniProfiler
- Flux chamber incubations in situ - MiniChamber Lander
- Non invasive flux measurements - Eddy Covariance
- Deep sea landers
- Measurements in the water column
- Working with very low O₂ concentrations - STOX sensor

Unisense Field Systems



Microsensors for in situ use

- Any Unisense sensor for in situ use:
 - O₂, H₂S, H₂, N₂O, NO
 - pH, Redox potential, electrical potential
- Tip size: 10 - 500 µm
- Mounted via In Situ Connector
- Pressure compensation (> 50 m)



Lab and Field Multi Meter



Deep sea: 6000 m water depth



Shallow: 300 m water depth

Application of microsensors

Microprofiles:

- Small tip diameter
- High spatial resolution



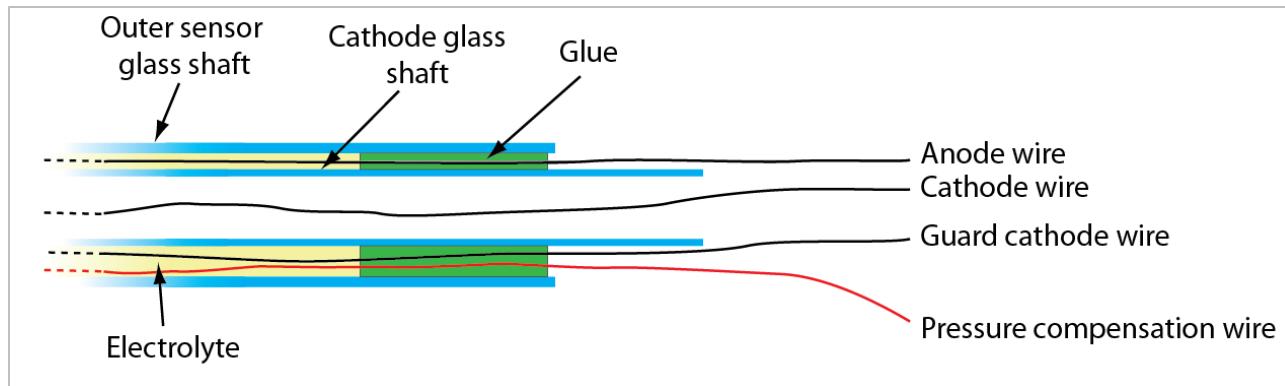
Bulk water:

- 500 μm tip diameter
- Protection cap



Pressure compensation

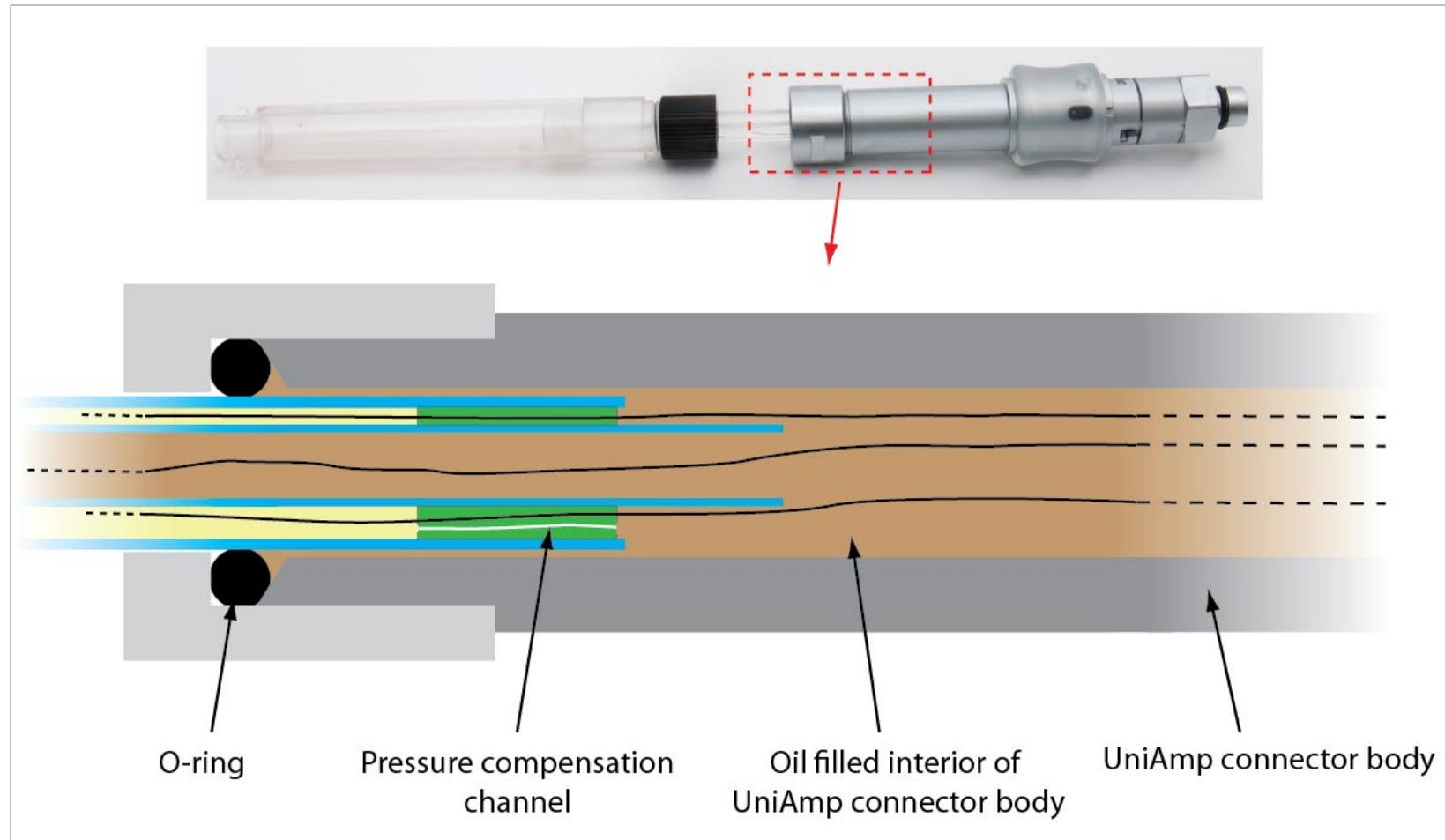
Pressure compensation is required for > 50 m water depth



In situ connectors

Pressure compensation

Sensor mounted in In Situ Connector



SensorTrace Suite



Logger (freeware)



Profile



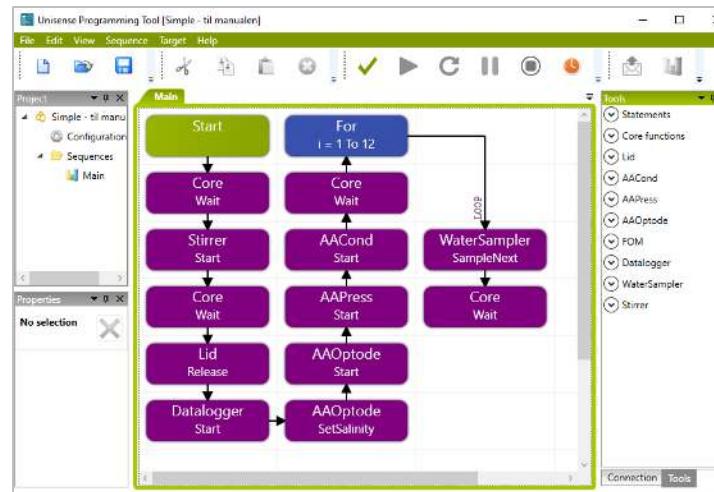
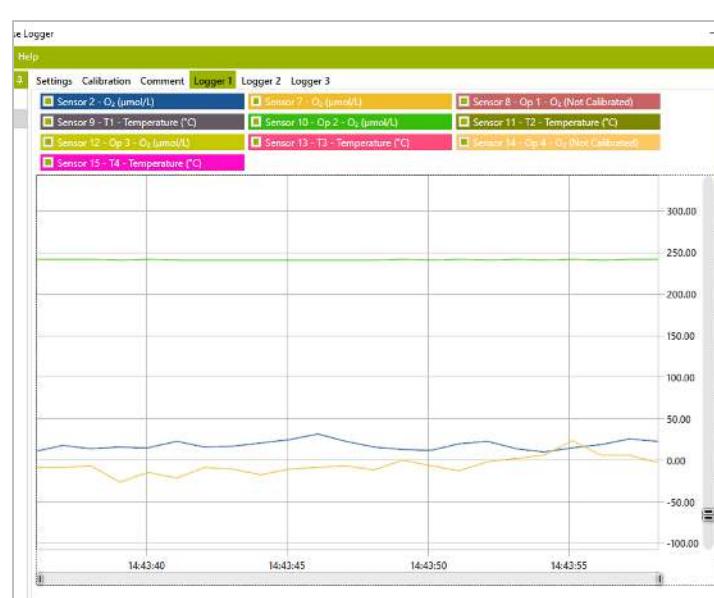
Rate



Photo



Programming tool

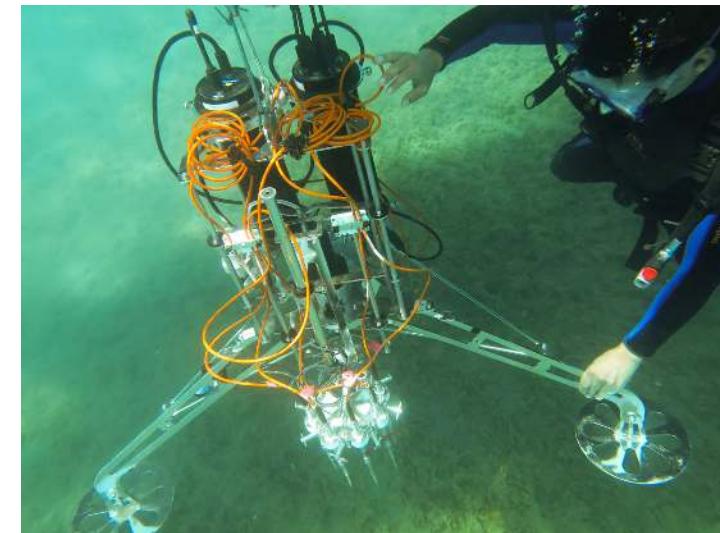


Profile
Logger (freeware)

Programming tool

In Situ MiniProfiler

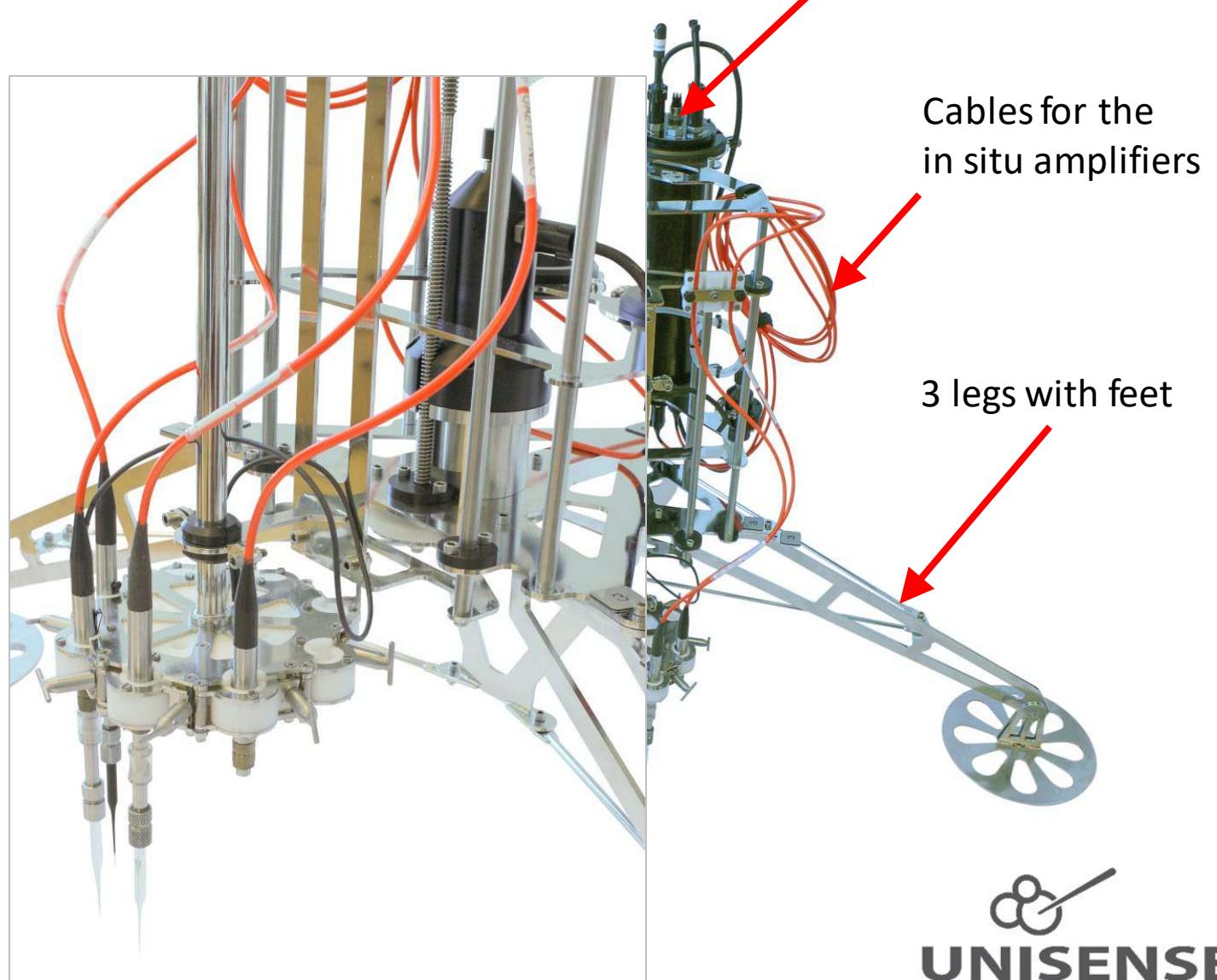
- Depth distribution of analytes, e.g.:
 - O₂ penetration
 - H₂S front
 - pH depth distribution
 - Electrical potential (cable bacteria)
- Fluxes in the analysed spot
- Specific activity per volume of sediment with depth
- Activity of the sediment itself without macro fauna
- Diurnal cycles - non destructive, repeated measurements



Diver assisted deployment
of MiniProfiler

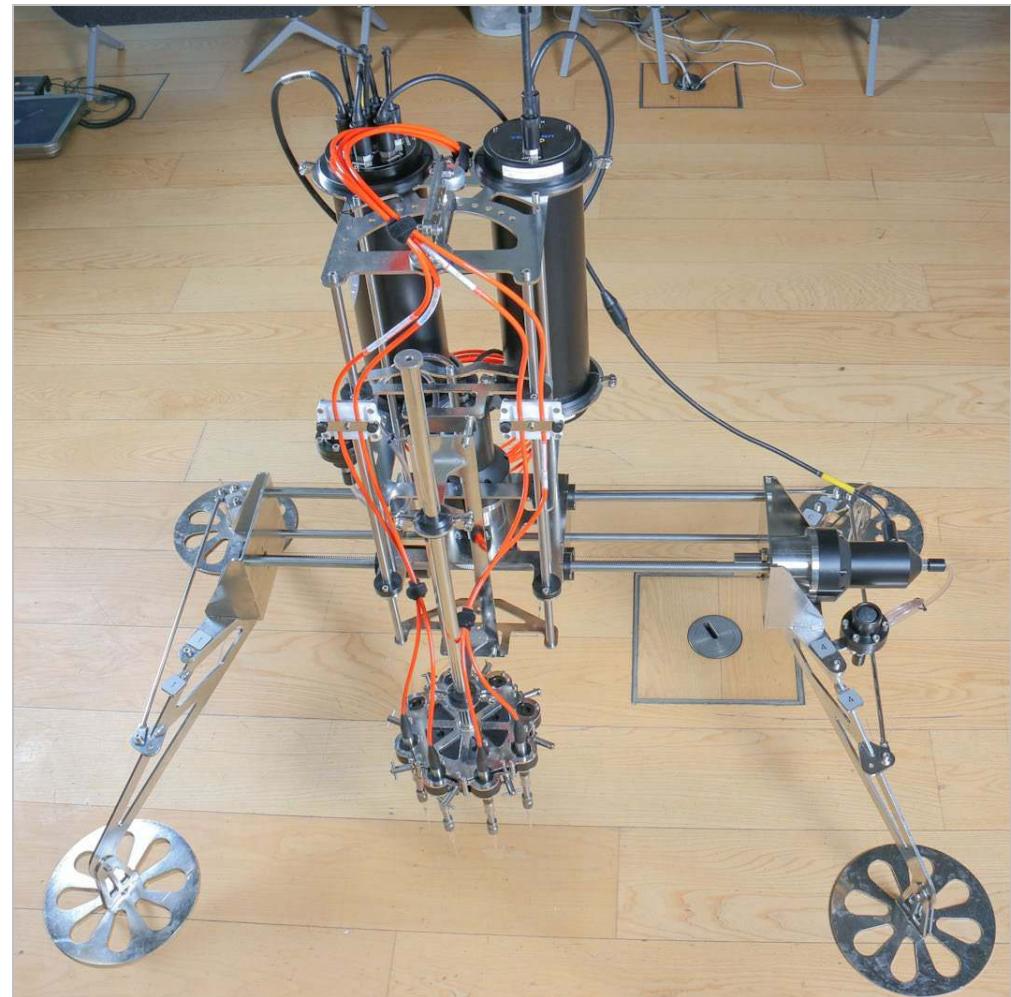
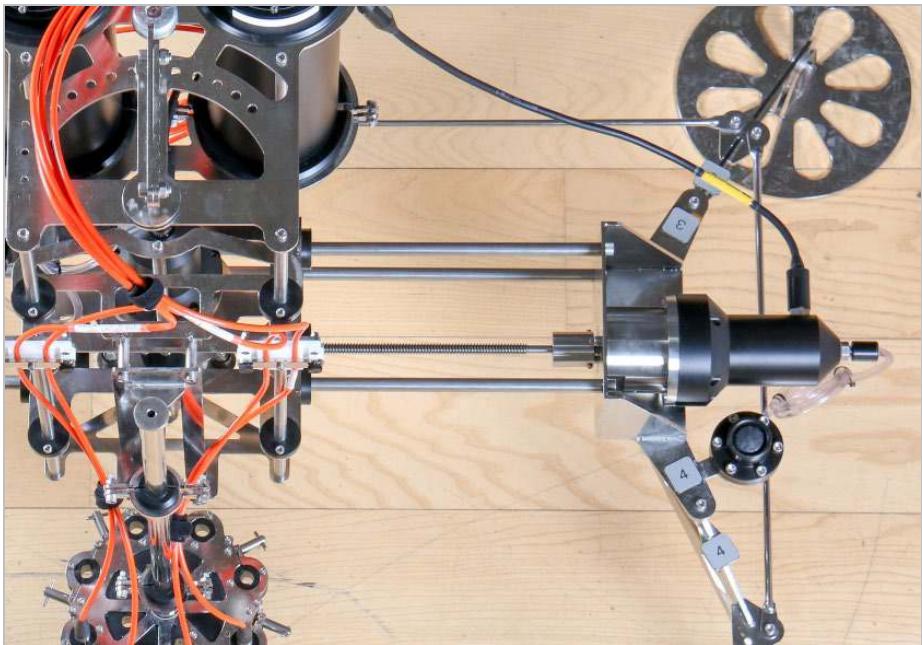
In Situ MiniProfiler – 1D

- Tripod
- 1.5 m between feet
- Controlled by the Field DataLogger
- Room for extra battery pack
- 4 or 8 sensor version
- 300 or 6000 m depth rating
- 50 μm step size



In Situ MiniProfiler – 2D

- Vertical unit same as 1D
- Horizontal movement
- Microprofiles on a line

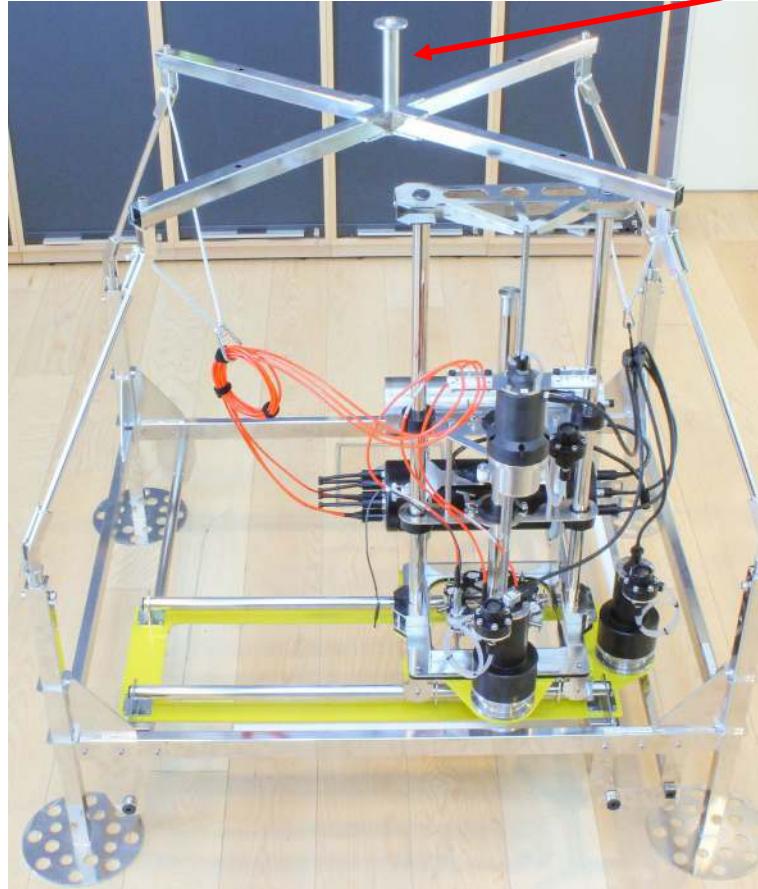


In Situ MiniProfiler – 3D

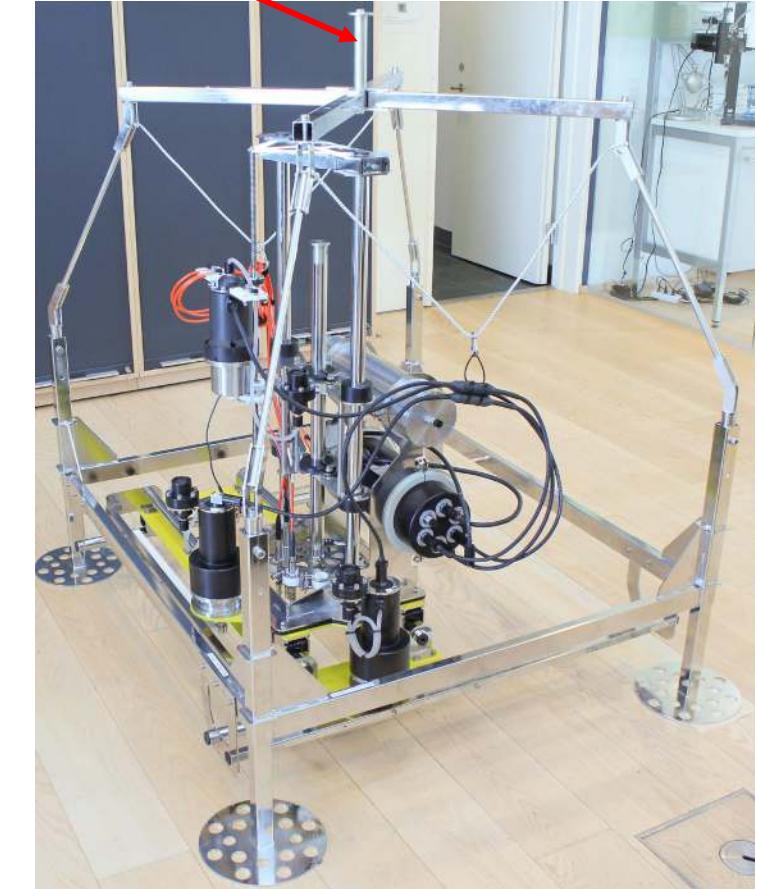
- Vertical and horizontal movement
- Microprofiles in a grid



ROV Switch

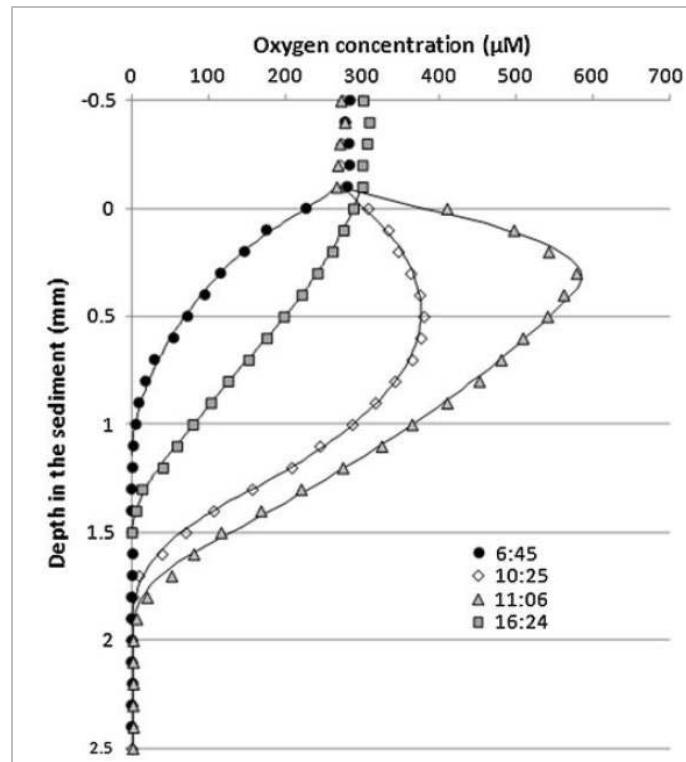


ROV Handle

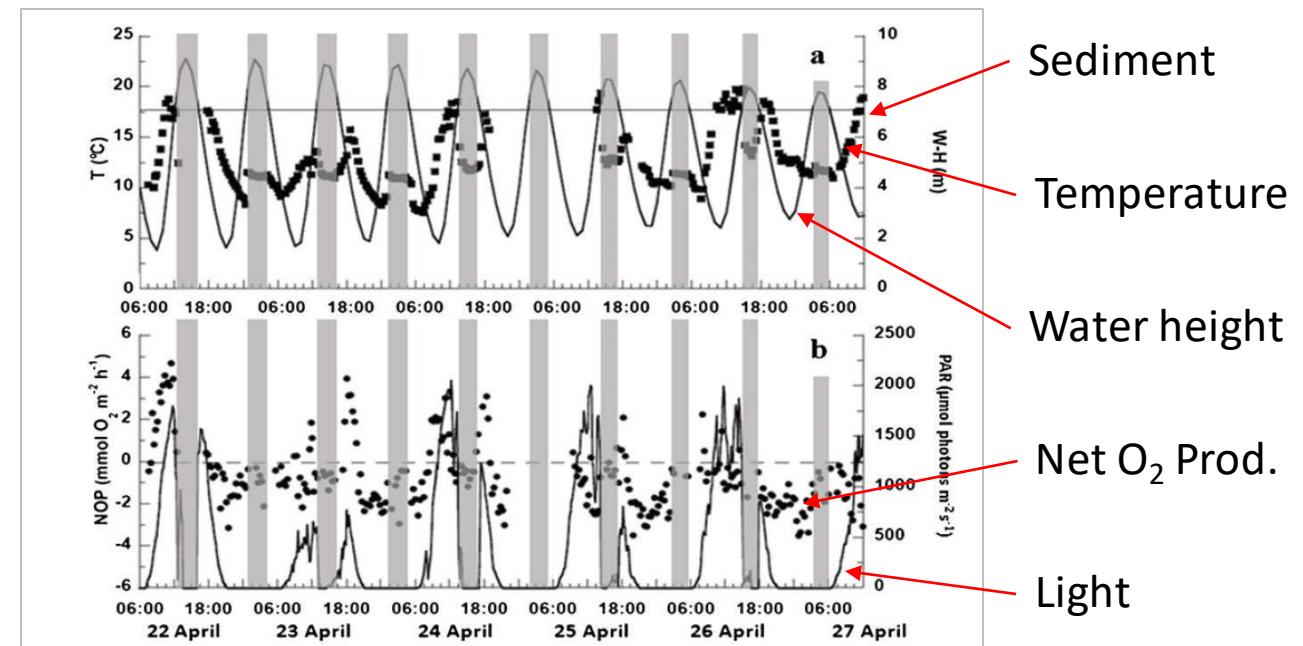


In Situ MiniProfiler – Examples of data

Oxygen dynamics on intertidal flat in the English Channel, France
6-day experiment, > 900 O₂ microprofiles



Examples of microprofiles

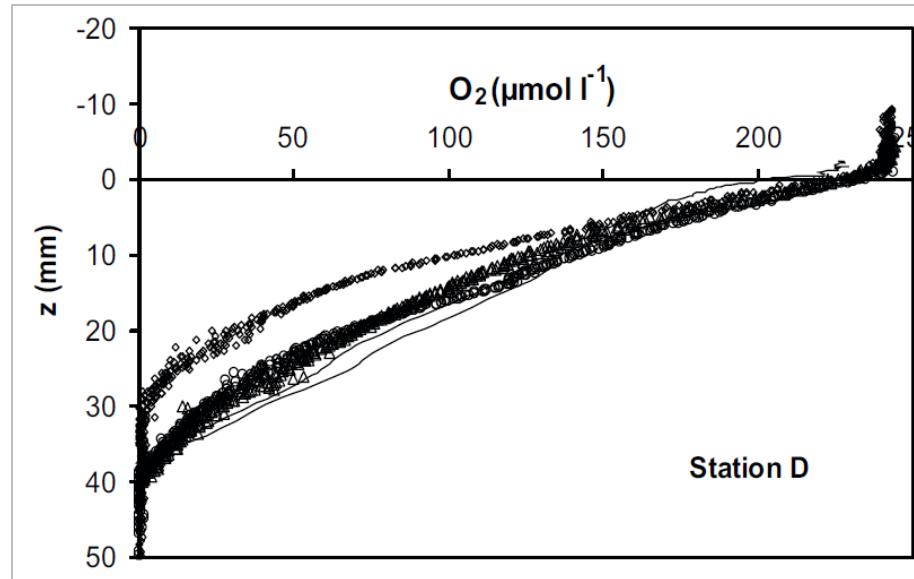


Net Oxygen Production (NOP), light (PAR),
temperature and water height

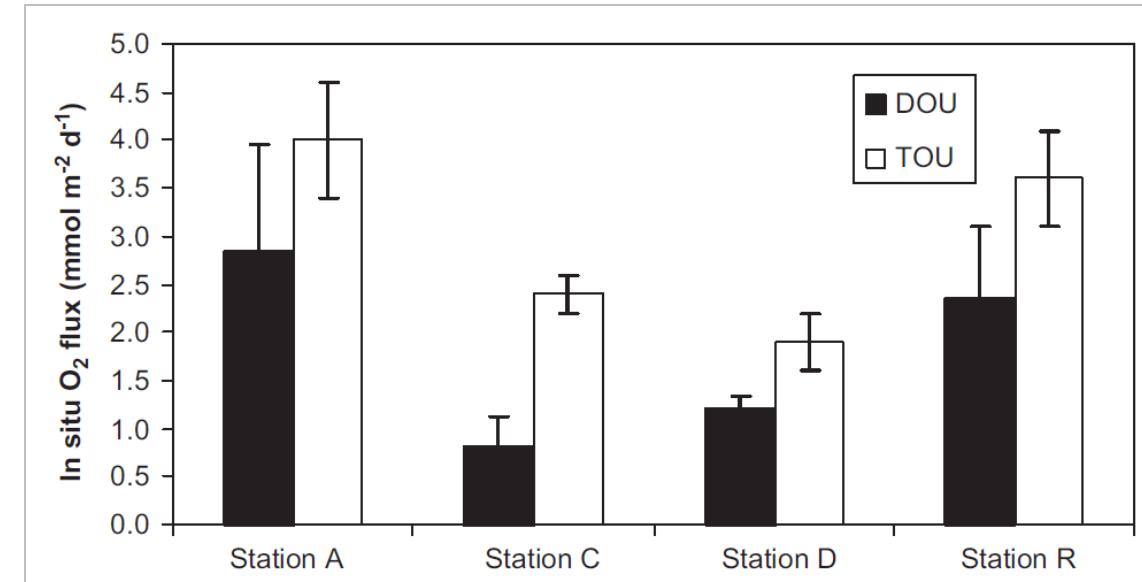
Denis, L., F. Gevaert, and N. Spilmont. 2012. Microphytobenthic production estimated by in situ oxygen microprofiling: Short-term dynamics and carbon budget implications. *J. Soils Sediments* 12: 1517–1529. doi:10.1007/s11368-012-0588-8

In Situ MiniProfiler – Examples of data

Oxygen penetration and uptake rates in the Southern Atlantic off Congo
Autonomous lander, 1304 – 3994 m water depth



Station D at 3964 m

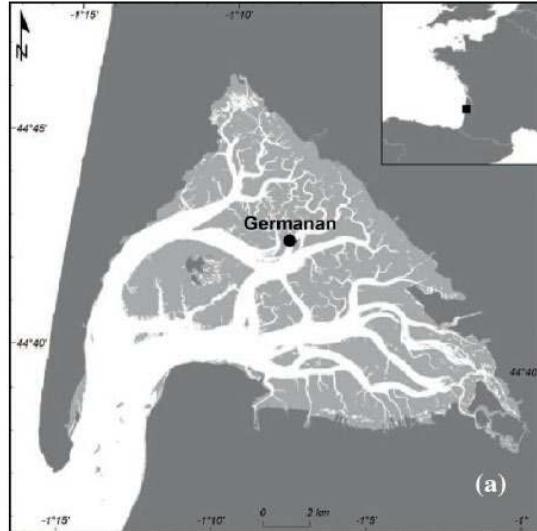


DOU: Diffusive O_2 uptake from microprofiles

TOU: Total O_2 uptake from flux chamber incubation

Rabouille, C., J. C. Caprais, B. Lansard, P. Crassous, K. Dedieu, J. L. Reyss, and A. Khripounoff.
2009. Organic matter budget in the Southeast Atlantic continental margin close to the Congo
Canyon: In situ measurements of sediment oxygen consumption. Deep. Res. Part II Top. Stud.
Oceanogr. 56: 2223–2238. doi:10.1016/j.dsr2.2009.04.005

In Situ MiniProfiler – Examples of data

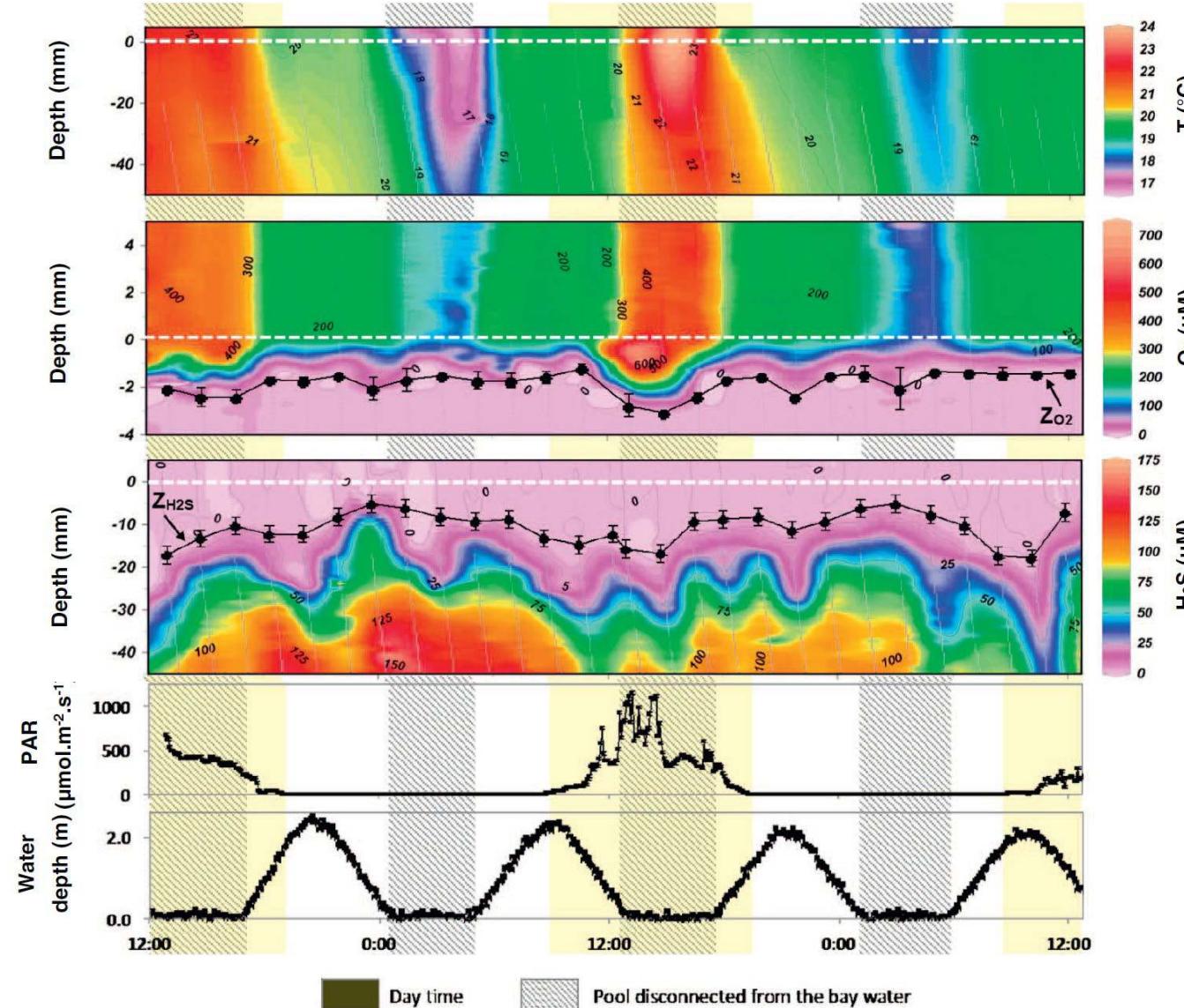


Microprofiles in tidal pools
Arcachon Bay (France)

- Tidal cycles - 2 m
- Temperature
- O₂
- H₂S

Rigaud, S., B. Deflandre, O. Maire, G. Bernard, J. C. Duchêne, D. Poirier, and P. Anschutz. 2018. Transient biogeochemistry in intertidal sediments: New insights from tidal pools in *Zostera noltei* meadows of Arcachon Bay (France). Mar. Chem. 200: 1–13. doi:10.1016/j.marchem.2018.02.002

In Situ MiniProfiler – Examples of data



Rigaud, S., B. Deflandre, O. Maire, G. Bernard, J. C. Duchêne, D. Poirier, and P. Anschutz. 2018. Transient biogeochemistry in intertidal sediments: New insights from tidal pools in *Zostera noltei* meadows of Arcachon Bay (France). Mar. Chem. 200: 1–13.
doi:10.1016/j.marchem.2018.02.002



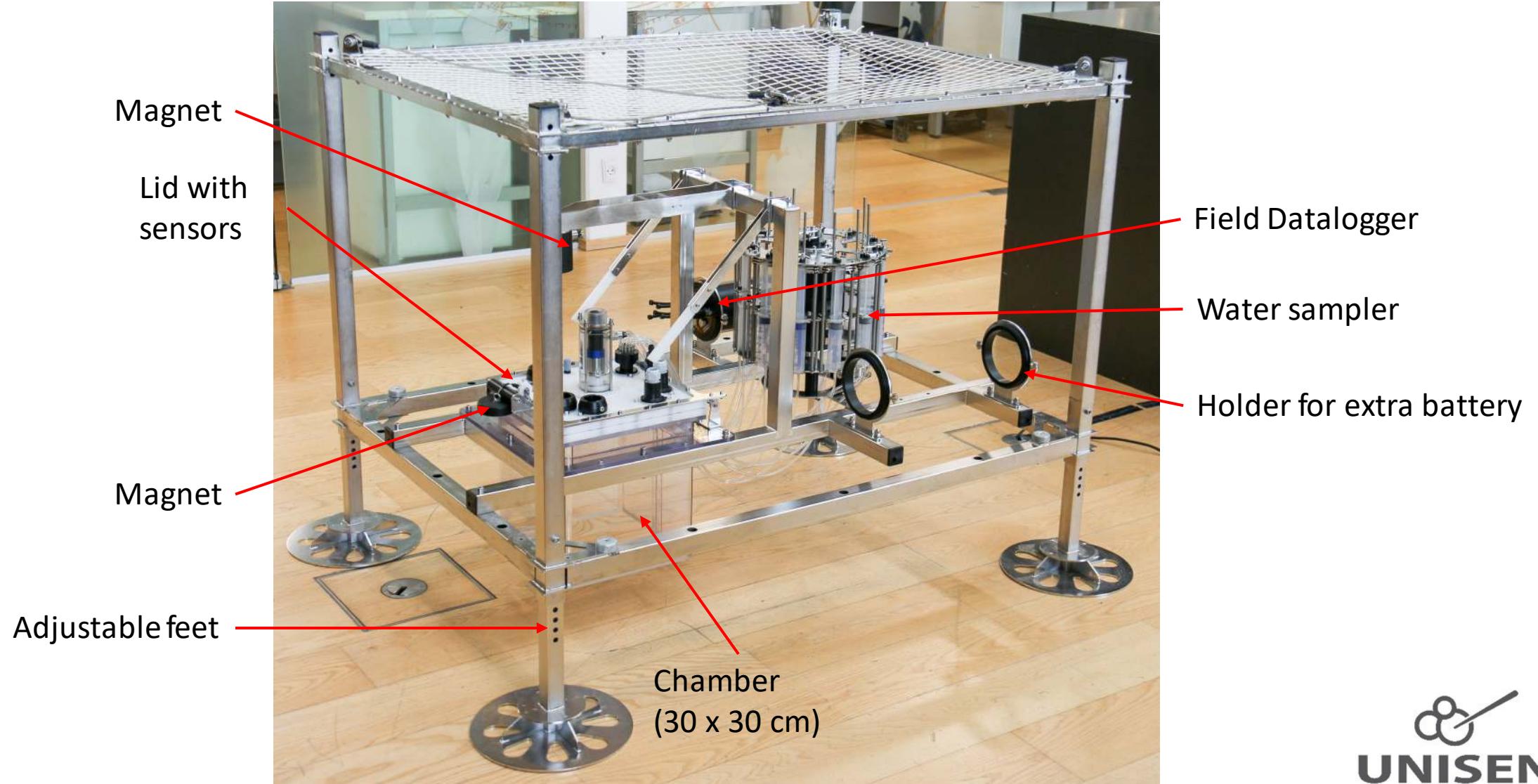
UNISENSE

MiniChamber Lander

- Sediment-water fluxes
- Use sensors
 - Unisense sensors
 - Aanderaa sensors (O_2 , conductivity, depth, turbidity)
- Use water samples
 - 12 samples during incubation
 - 60 ml sample for any analysis
- Flux estimates of larger sediment area
 - Includes activity of flora and fauna
- Ratio of flux from chamber to diffusive flux indicate faunal activity



MiniChamber Lander

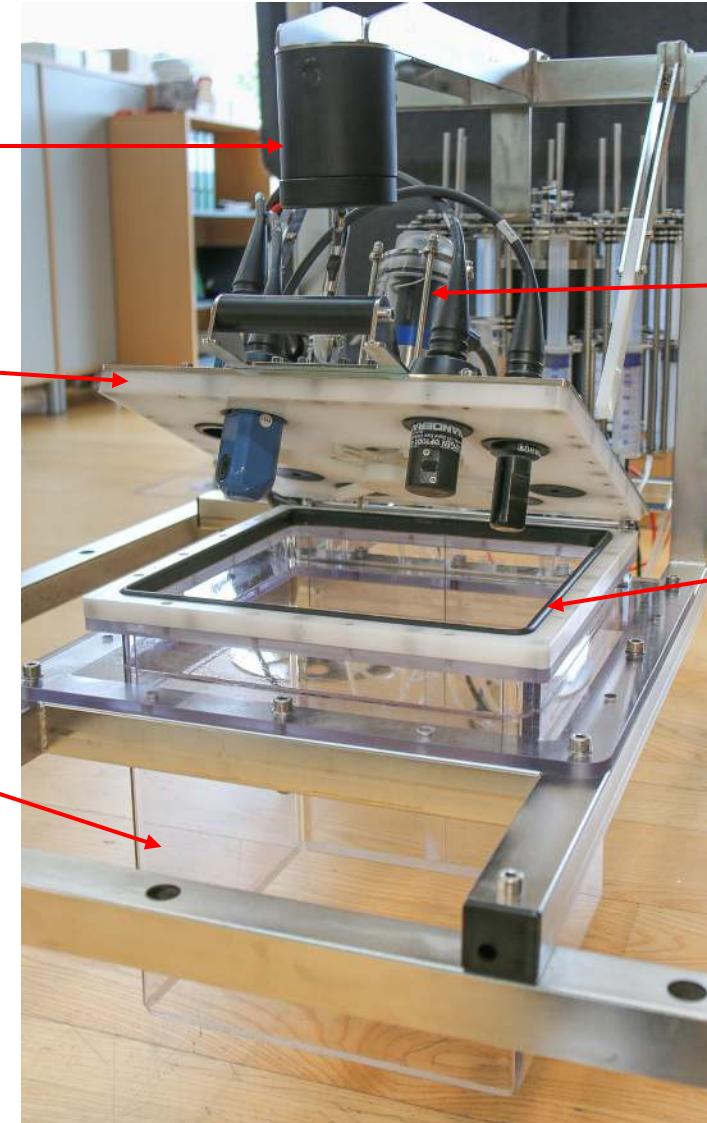


Chamber unit and lid

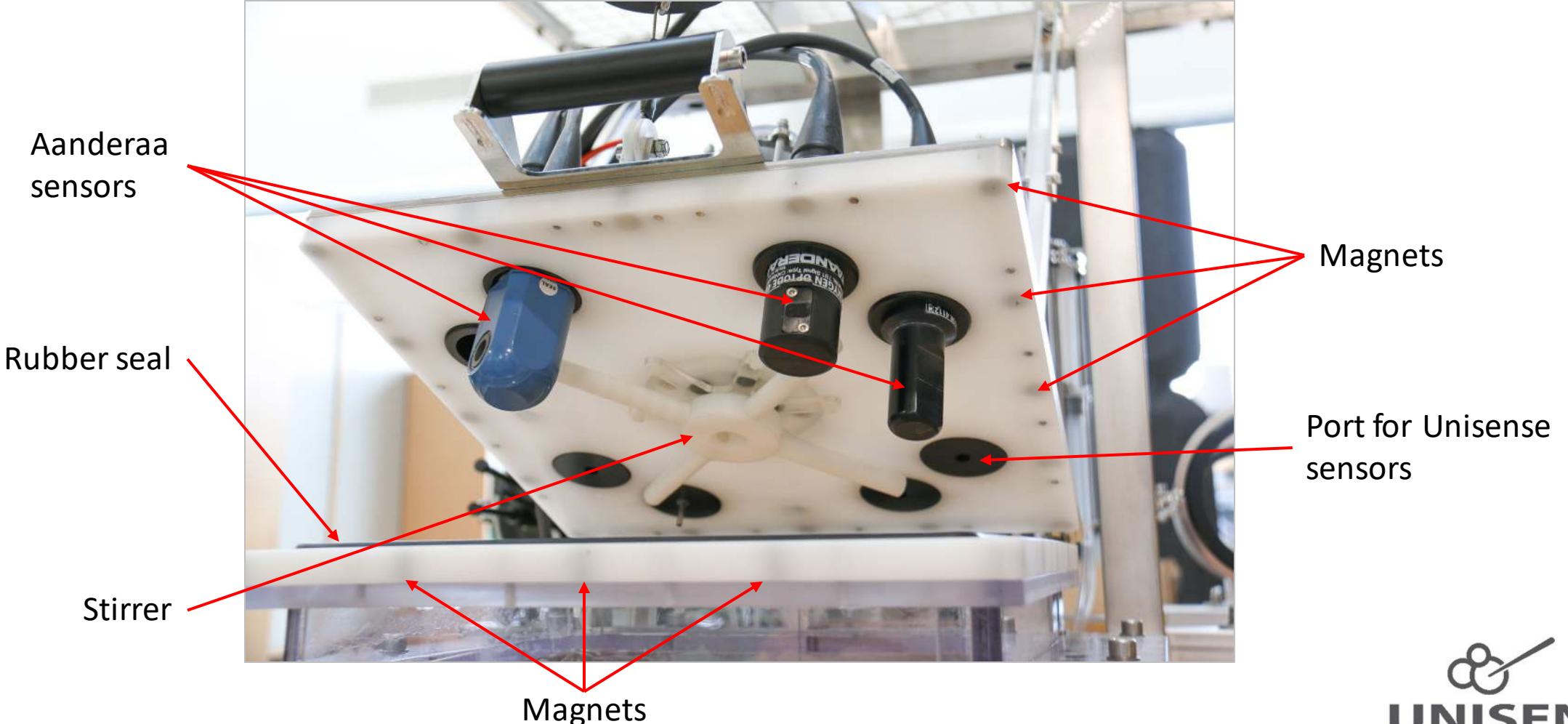
Permanent magnet &
counter polarized electromagnet

Lid (POM, Polyoxymethylene)

Incubation chamber (Polycarbonate)

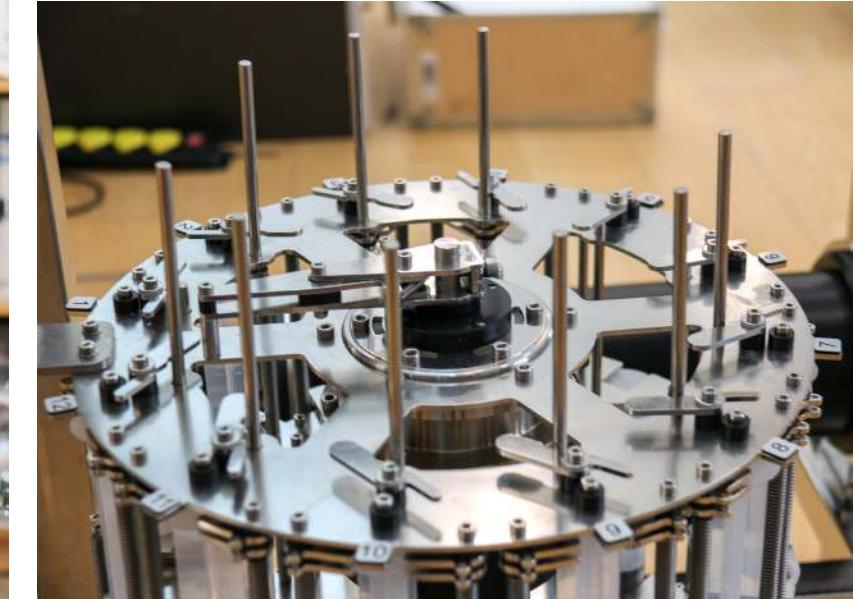


Lid - below



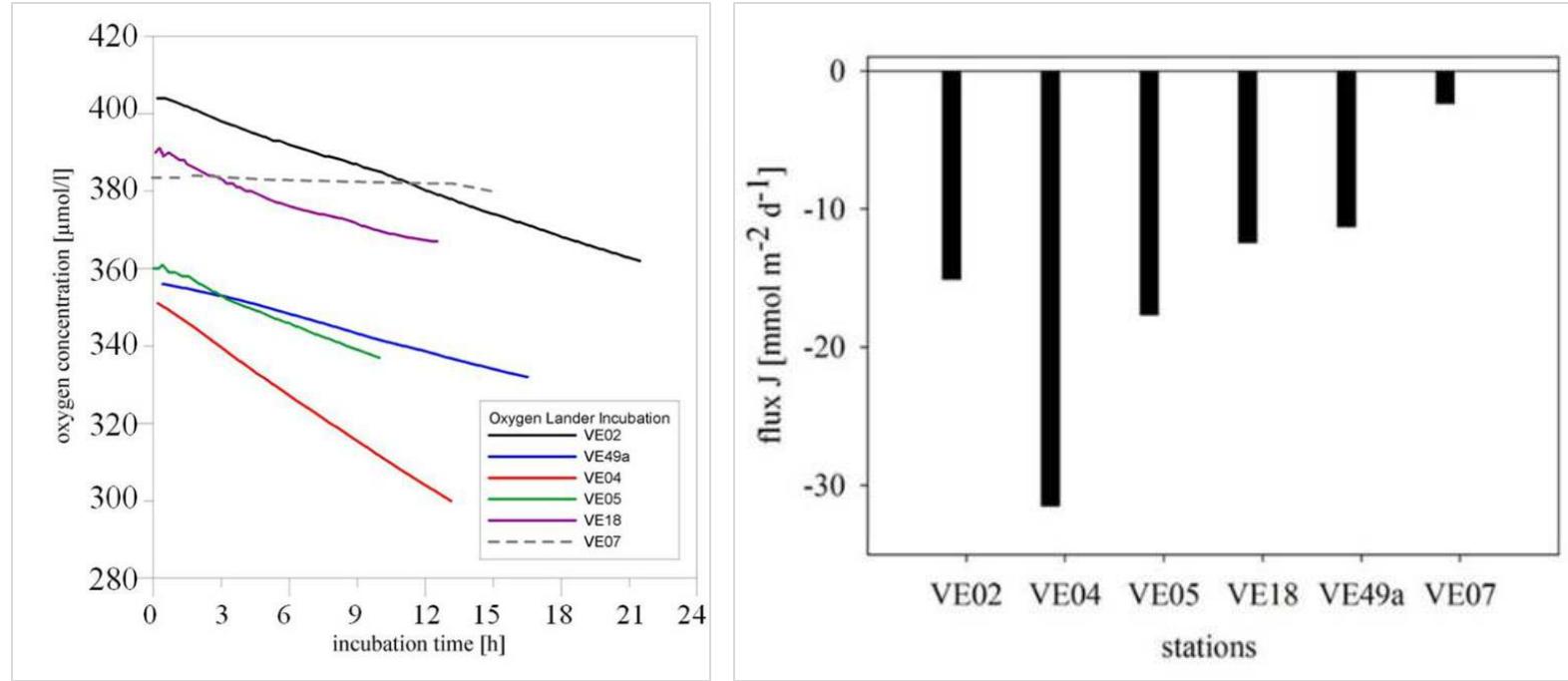
Water sampler

- 12 syringes
- 60 ml
- Sample or inject
- Quantify sediment-water fluxes of e.g. nutrients



Examples of data

Oxygen uptake rates at different stations in the Baltic Sea



Deployment data

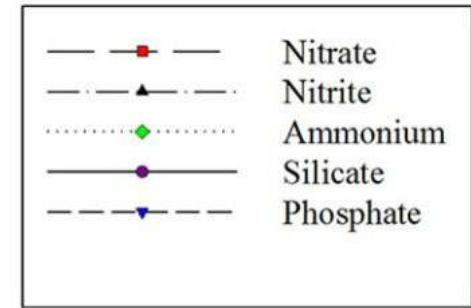
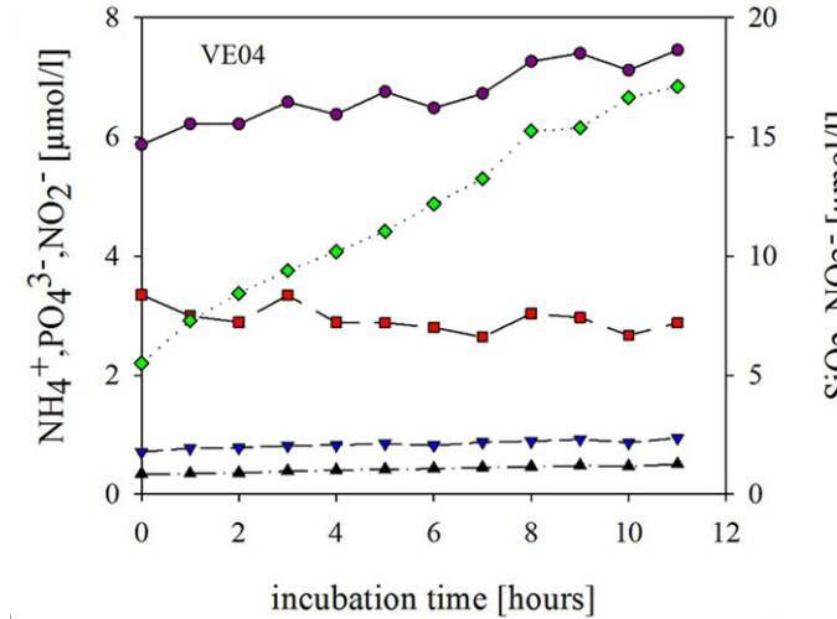
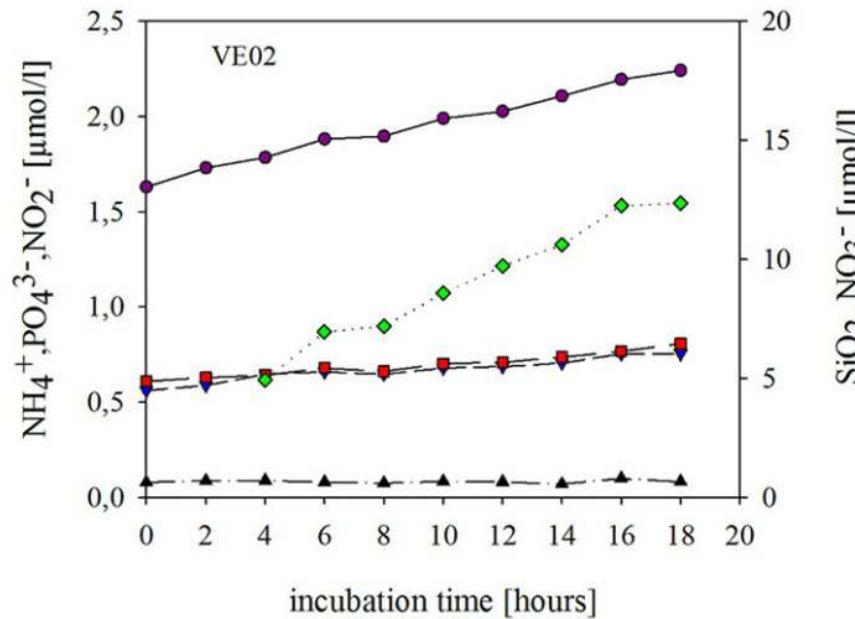
- Chamber penetration into sediment: 5 cm
- Chamber water volume: 27 l
- Incubated sediment area: 900 cm²
- Stirring: 15 rpm
- Incubation length: 10 - 22 hours

MiniChamber Lander with O₂ optode and water sampler

Thoms, F., C. Burmeister, J. W. Dippner, M. Gogina, U. Janas, H. Kendzierska, I. Liskow, and M. Voss. 2018. Impact of macrofaunal communities on the coastal filter function in the Bay of Gdańsk, Baltic Sea. *Front. Mar. Sci.* 5: Article no. 205.

Examples of data

Sediment water fluxes of nutrients at different stations in the Baltic Sea

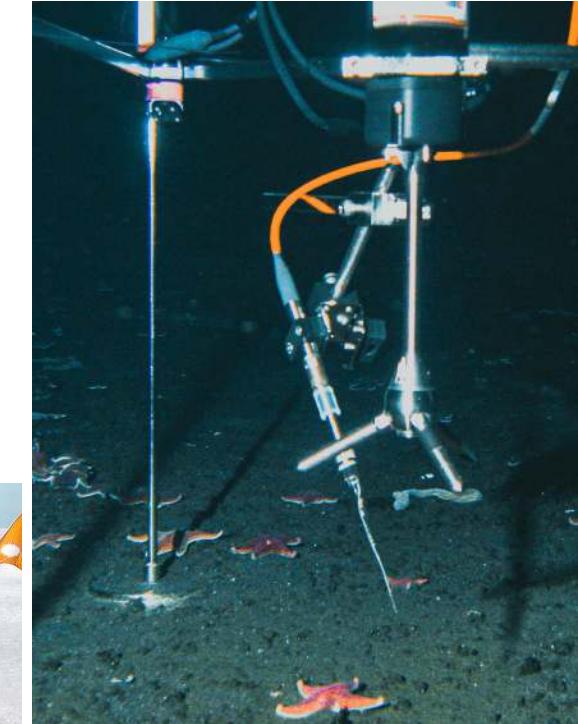


MiniChamber Lander with O_2 optode and water sampler

Thoms, F., C. Burmeister, J. W. Dippner, M. Gogina, U. Janas, H. Kendzierska, I. Liskow, and M. Voss. 2018. Impact of macrofaunal communities on the coastal filter function in the Bay of Gdansk, Baltic Sea. *Front. Mar. Sci.* 5: Article no. 205.

Eddy Covariance Method

- Sediment-water fluxes
- Non-invasive
- Sensors
 - O₂ microsensor (fast)
 - O₂ microoptode (fast)
 - H₂S microsensor (fast)
- Flux estimate from a large sediment area (5 - 100 m²)
- Includes large flora and fauna e.g. seagrass beds, mussel beds



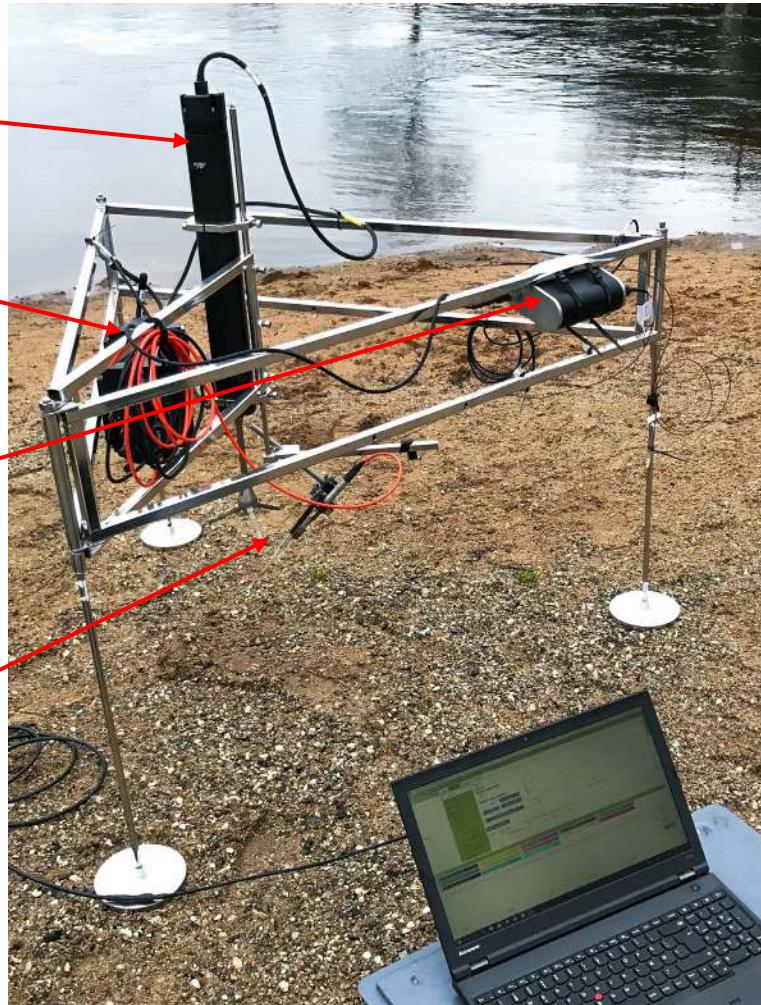
Eddy Covariance Method

Vector (acoustic 3D current meter)

Field Datalogger

Field Optode Meter

Sensors



Field Datalogger

Vector

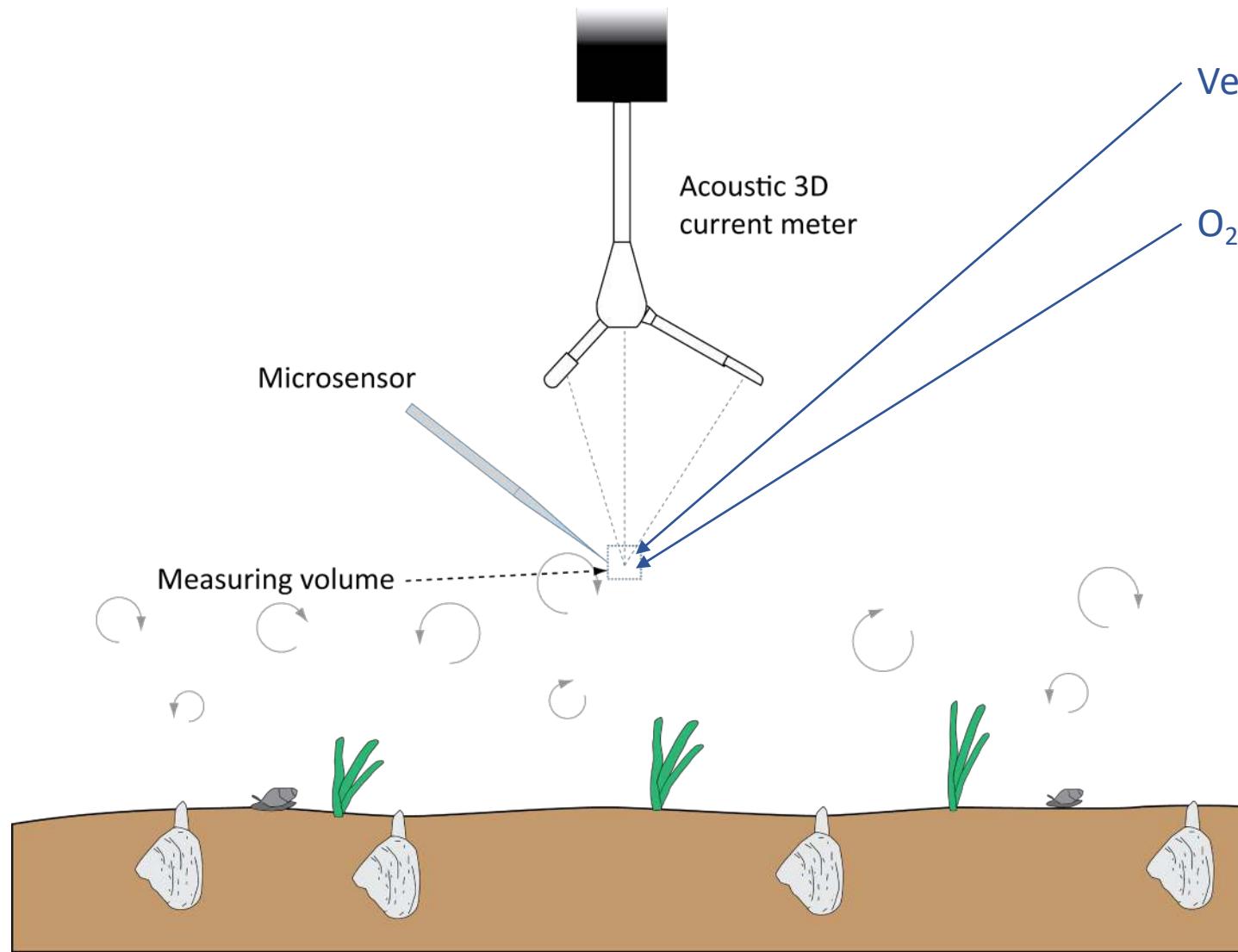
Clark type
 O_2 sensor

O_2 optode



UNISENSE

Eddy Covariance Method



Vertical flow rate (z)

64 Hz

O₂ concentration (C)

Principle of flux calculation

Instantaneous flux each time step (1/64 second)

$$\text{Flux} = z \times C$$

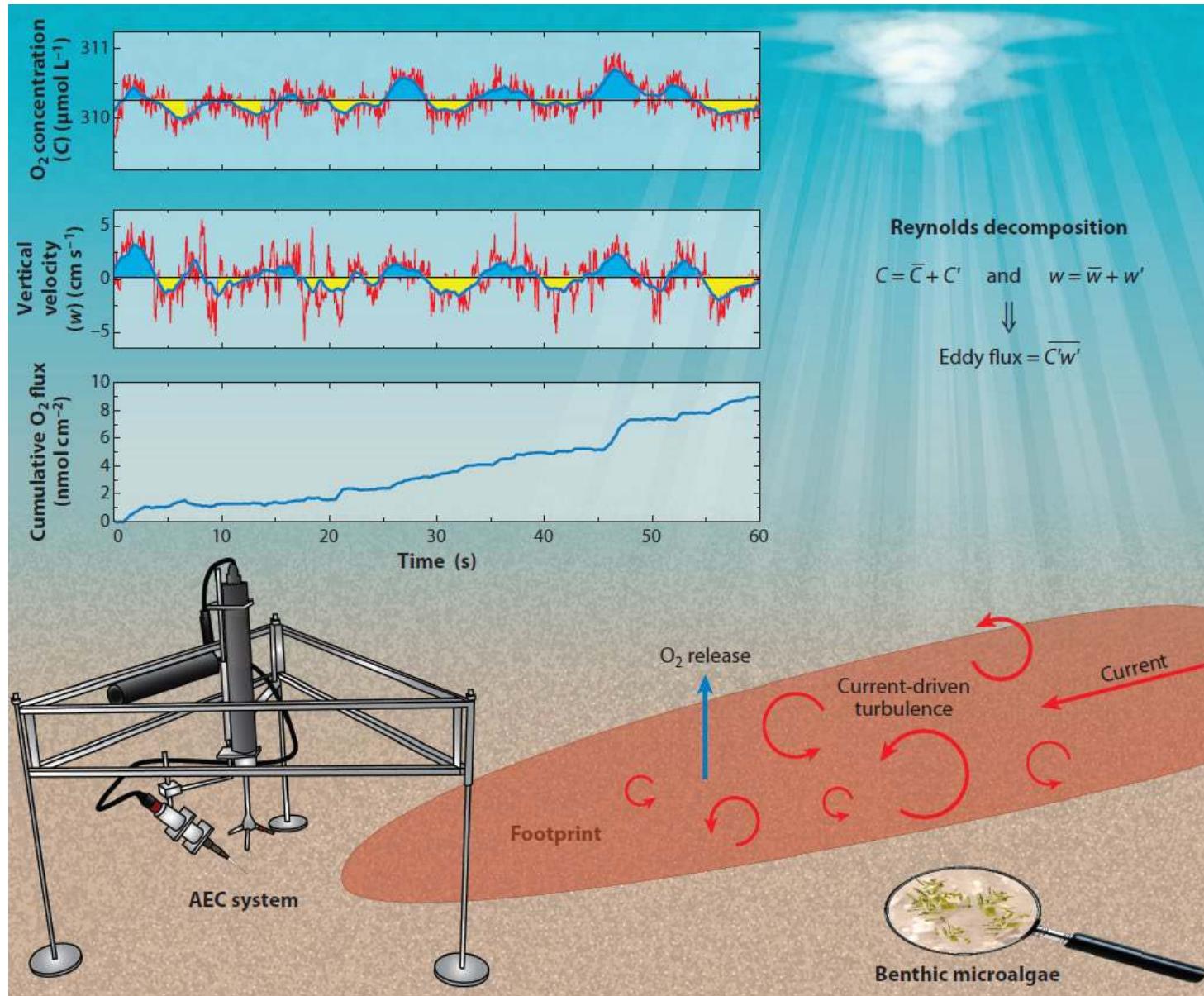
Summarize flux over time to get flux rates

Berg, P., M. L. Delgad, R. N. Glud, M. Huettel, C. E. Reimers, and M. L. Pace. 2017. Non-invasive flux Measurements at the Benthic Interface: The Aquatic Eddy Covariance Technique. Limnol. Oceanogr. e-Lectures 7: 1–50. doi:10.1002/loe2.10005



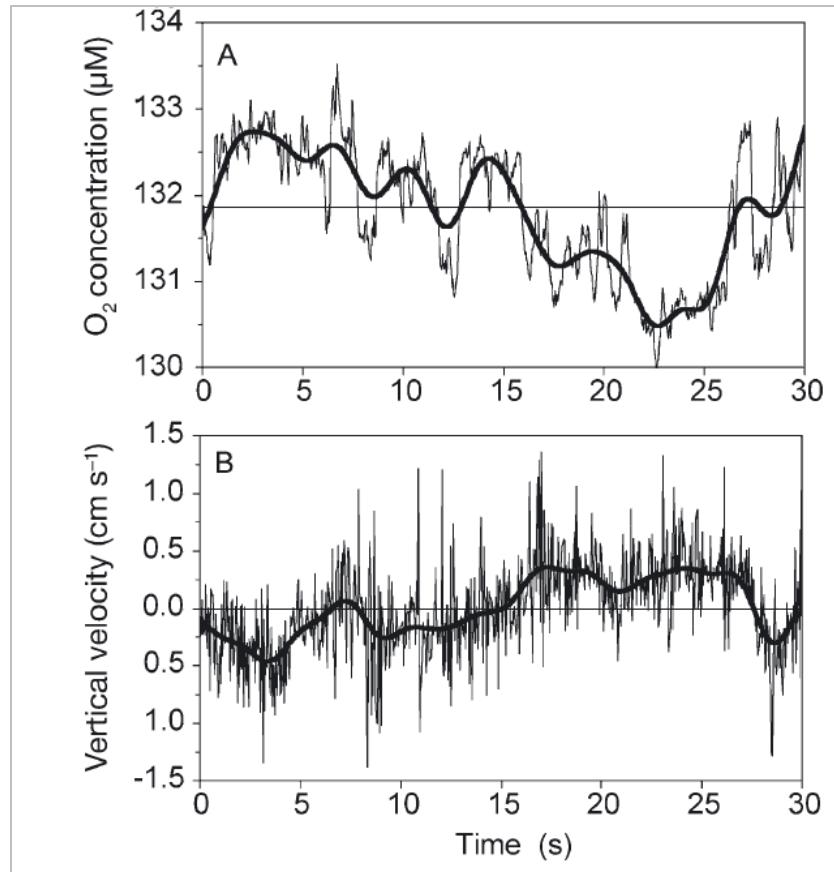
UNISENSE

Eddy Covariance - Principle

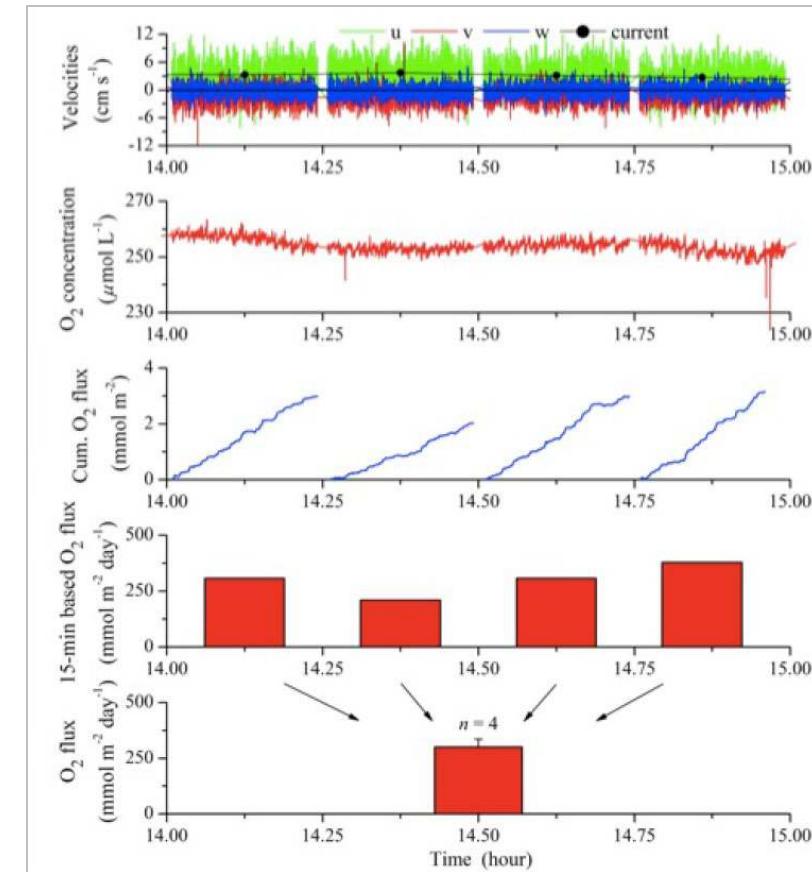


Berg, P., M. Huettel, R. N. Glud, C. E. Reimers, and K. M. Attard. 2022. Aquatic Eddy Covariance: The Method and Its Contributions to Defining Oxygen and Carbon Fluxes in Marine Environments. *Ann. Rev. Mar. Sci.* 14: 431–455.
doi:10.1146/annurev-marine-042121-012329

Eddy Covariance - Principle



Berg, P., H. Roy, F. Janssen, V. Meyer, B. B. Jørgensen, M. Huettel, and D. de Beer. 2003. Oxygen uptake by aquatic sediments measured with a novel non-invasive eddy-correlation technique. Mar. Ecol. Ser. 261: 75–83.

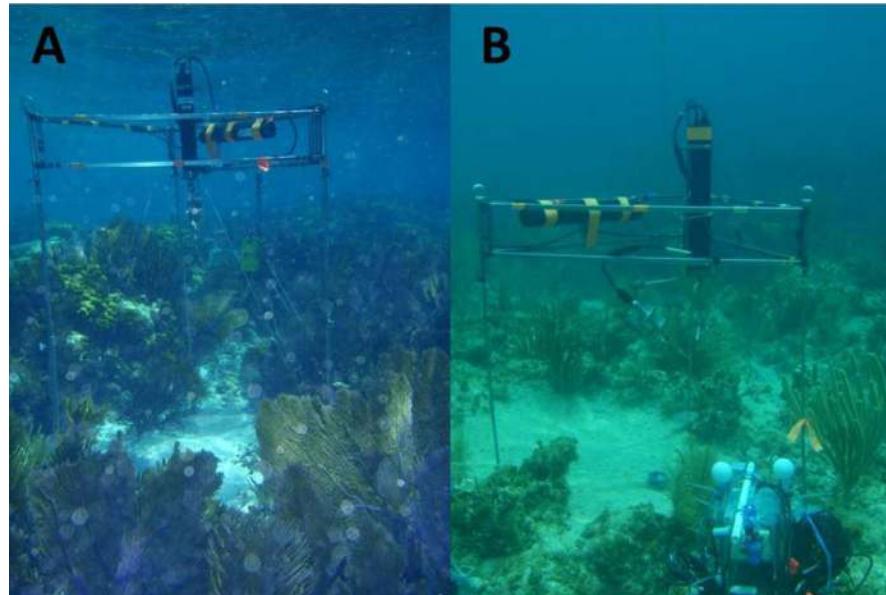


Berg, P., M. L. Delgard, R. N. Glud, M. Huettel, C. E. Reimers, and M. L. Pace. 2017. Non-invasive flux Measurements at the Benthic Interface: The Aquatic Eddy Covariance Technique. Limnol. Oceanogr. e-Lectures 7: 1–50. doi:10.1002/loe2.10005

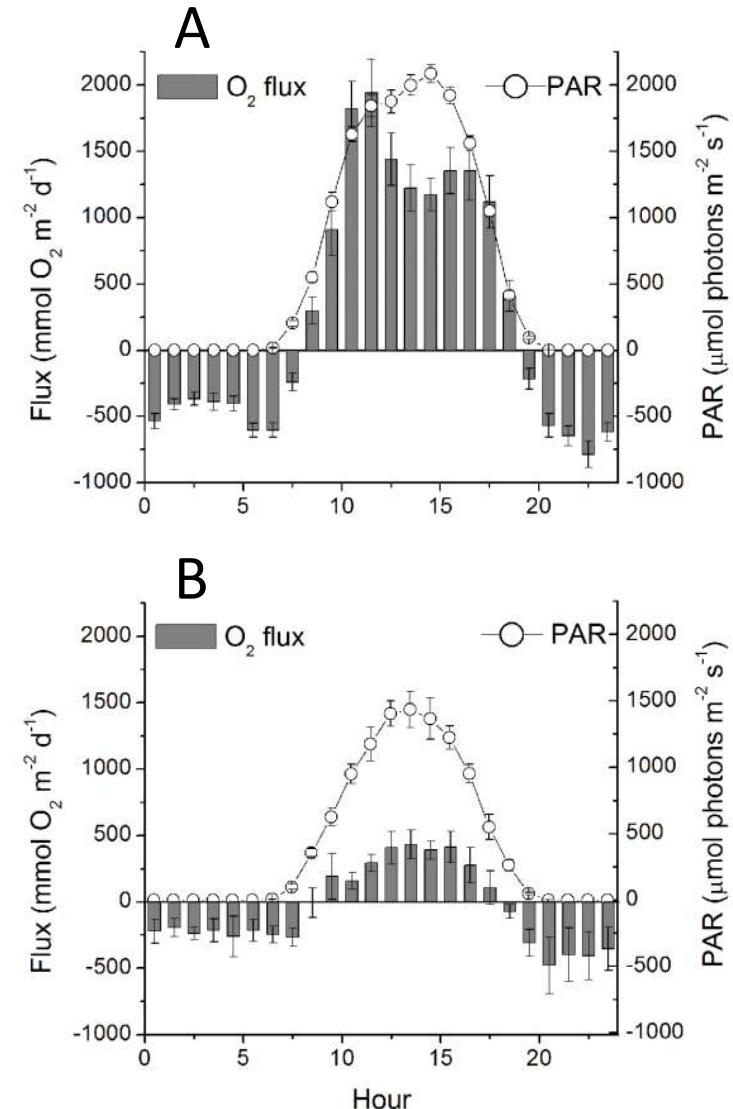


Eddy Covariance - Example of data

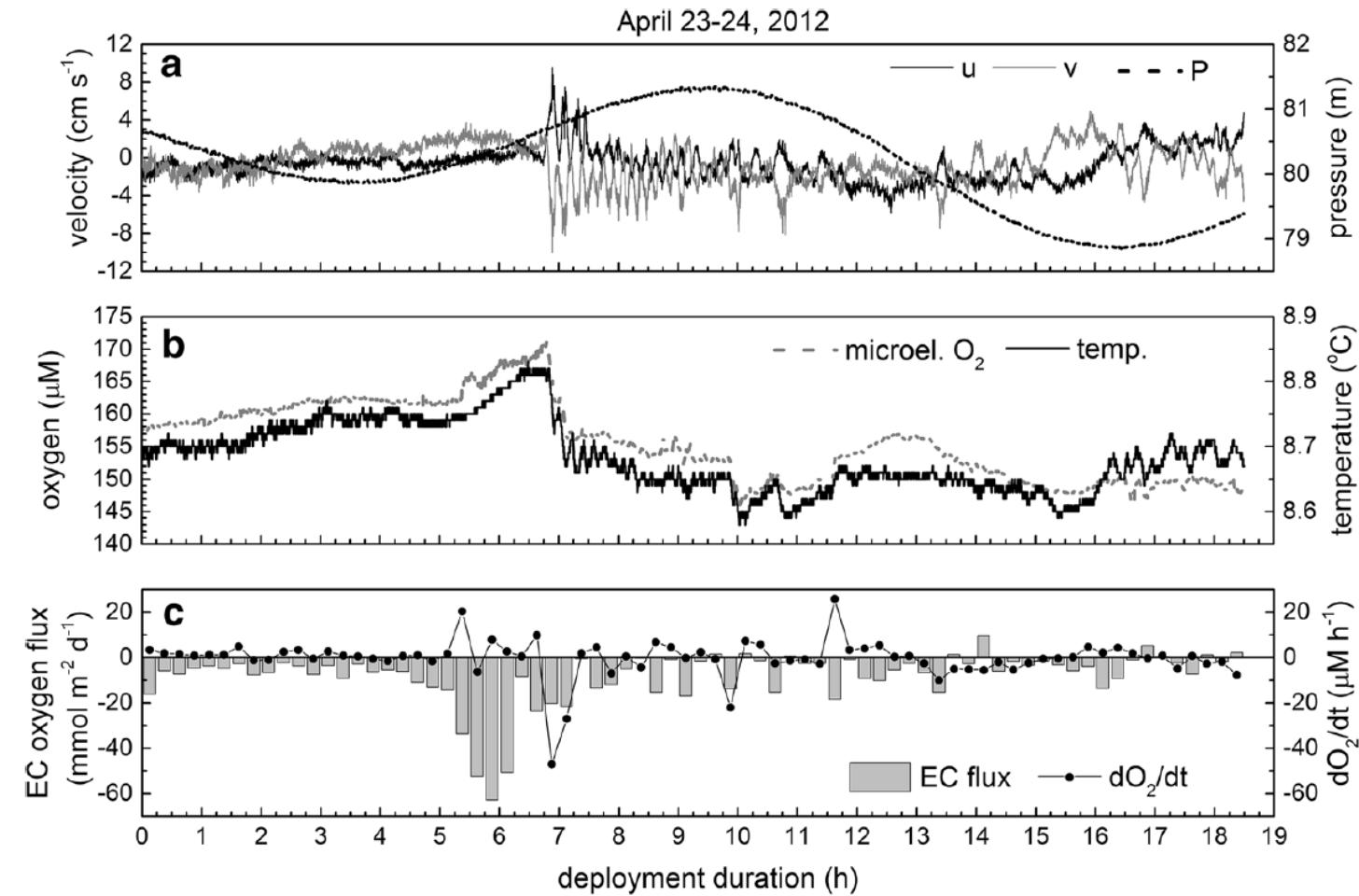
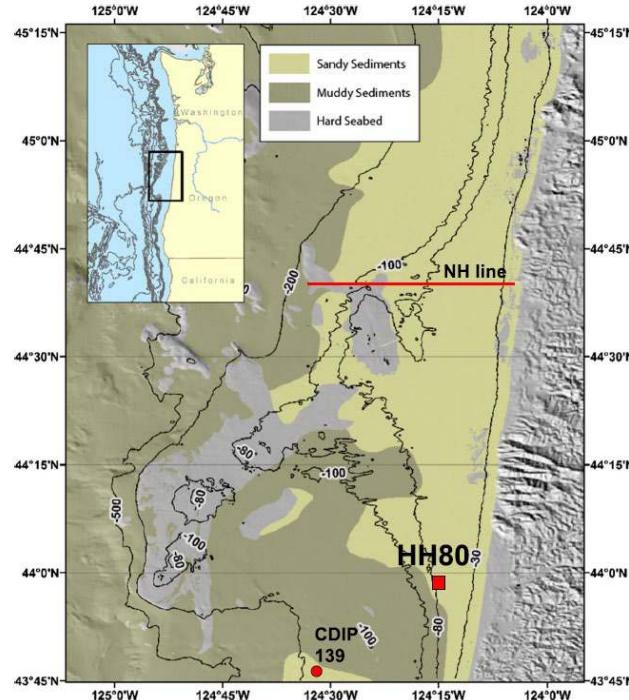
Coral reef – Oxygen fluxes



Long, M. H., P. Berg, D. de Beer, and J. C. Zieman. 2013.
In Situ Coral Reef Oxygen Metabolism: An Eddy
Correlation Study. PLoS One 8.
doi:10.1371/journal.pone.0058581



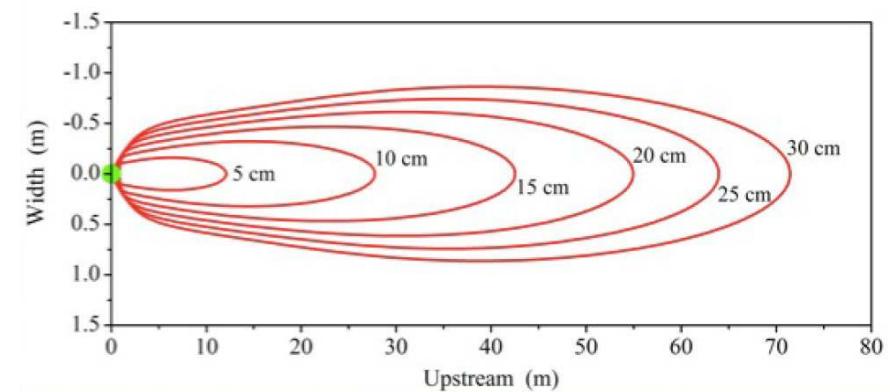
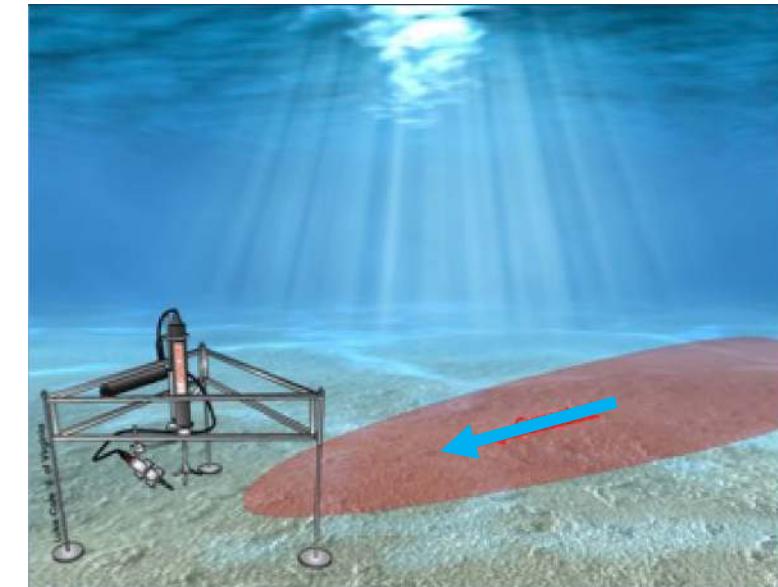
Eddy Covariance - Example of data



Reimers, C. E., H. T. Ozkan-Haller, R. D. Sanders, K. McCann-Grosvenor, P. J. Chace, and S. A. Crowe.
2016. The Dynamics of Benthic Respiration at a Mid-Shelf Station Off Oregon. *Aquat. Geochemistry* **22**: 505–527. doi:10.1007/s10498-016-9303-5

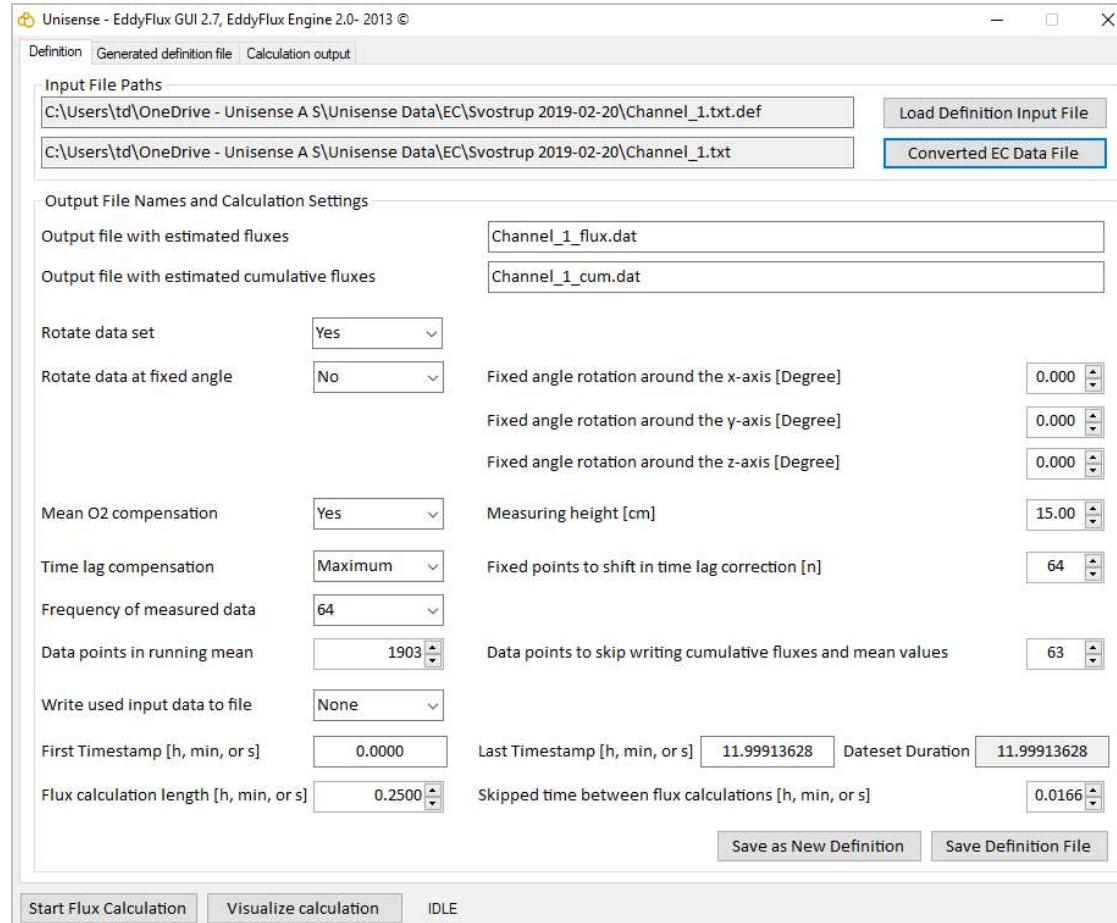
Eddy Covariance - Footprint

- Measured flux originates from downstream – The *footprint*
- Typical size 5 to 100 m²
- Calculated according to Berg et al., 2007
- Affected by
 - Sensor height over surface
 - Flow conditions
 - Sediment roughness



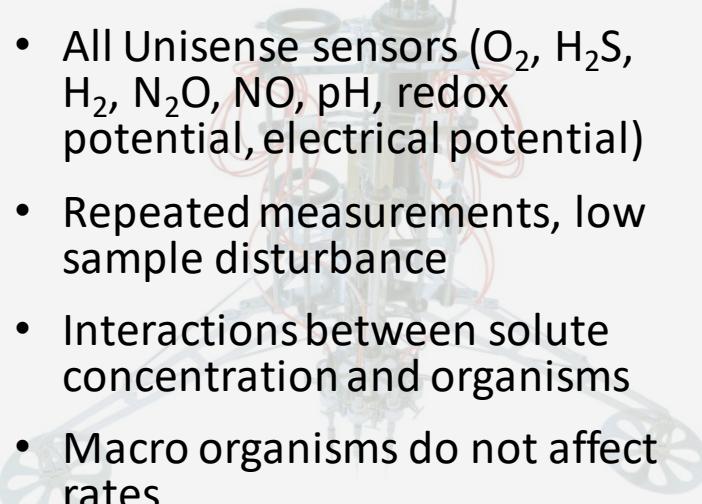
Berg, P., H. Roy, and P. L. Wiberg. 2007. Eddy correlation flux measurements: The sediment surface area that contributes to the flux. Limnol. Oceanogr. 52: 1672–1684.

Unisense EddyFlux Graphical User Interface

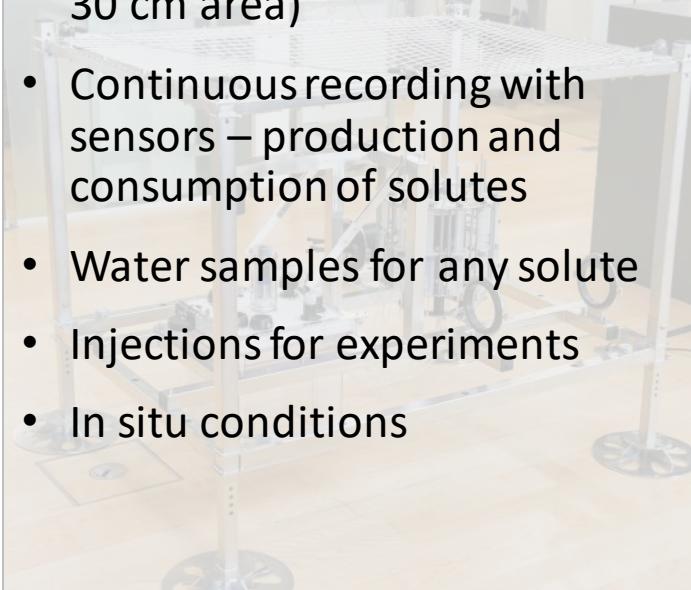


Three complementary methods

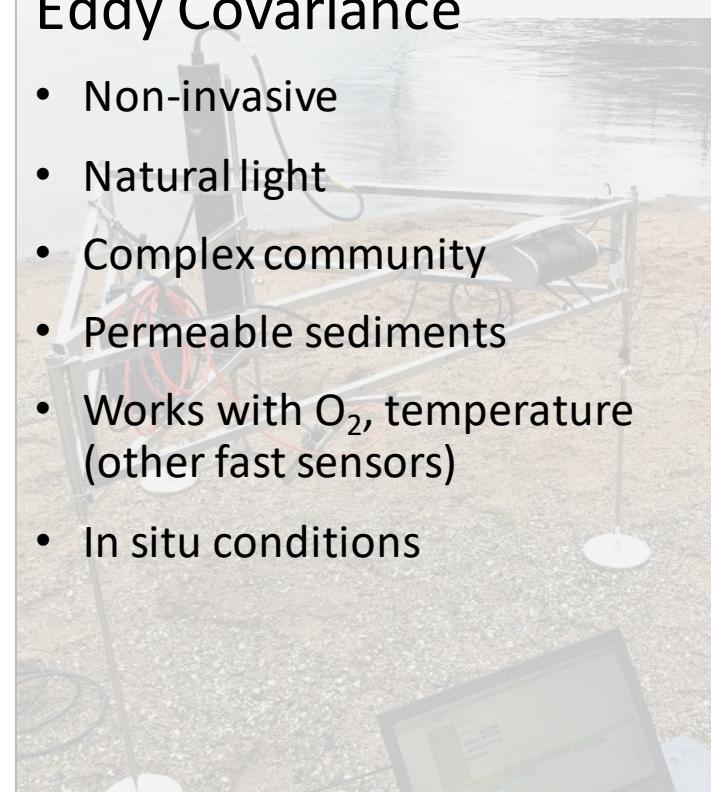
Microprofiles

- Vertical distribution of solutes
 - Consumption or production rates within the sediment
 - All Unisense sensors (O_2 , H_2S , H_2 , N_2O , NO , pH, redox potential, electrical potential)
 - Repeated measurements, low sample disturbance
 - Interactions between solute concentration and organisms
 - Macro organisms do not affect rates
 - In situ conditions
- 

MiniChamber

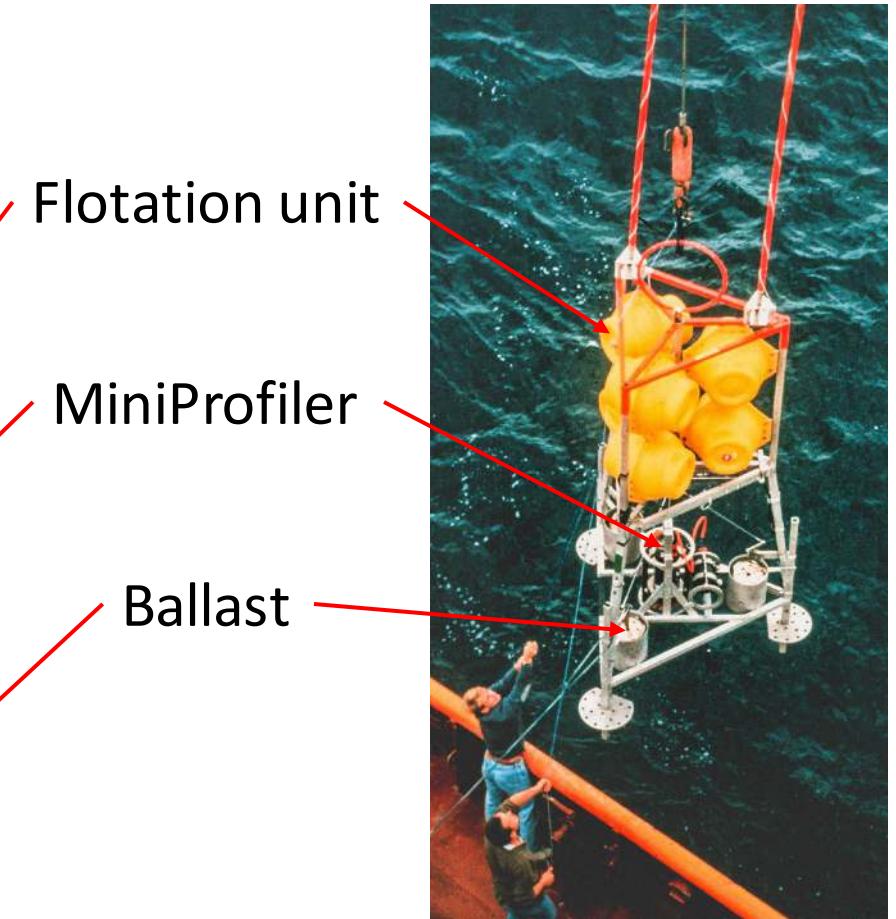
- Community metabolism (30 x 30 cm area)
 - Continuous recording with sensors – production and consumption of solutes
 - Water samples for any solute
 - Injections for experiments
 - In situ conditions
- 

Eddy Covariance

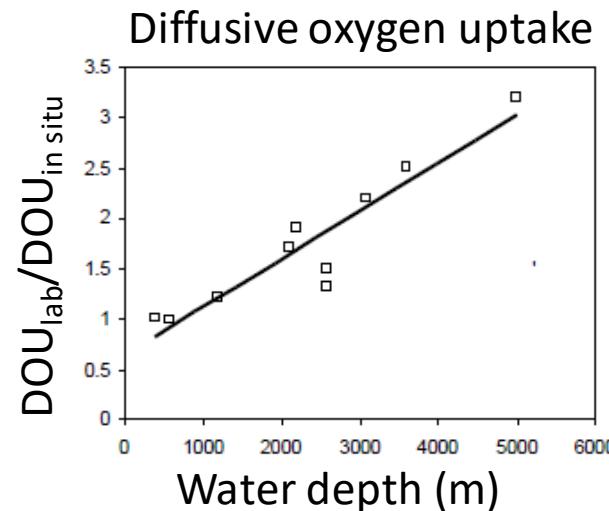
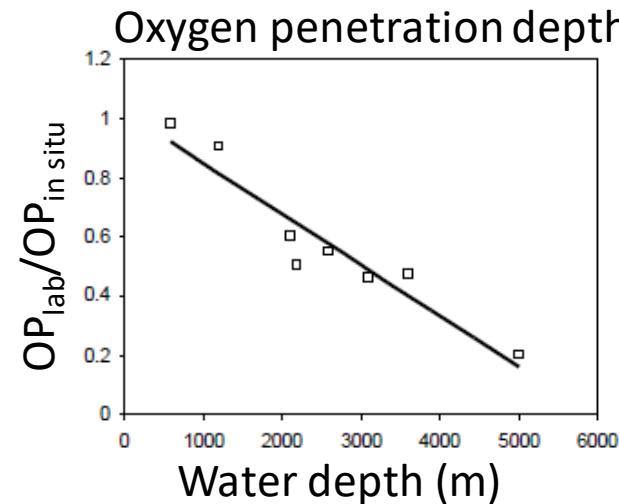
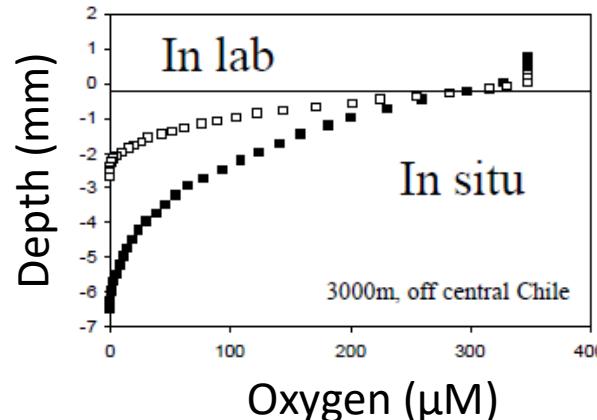
- Non-invasive
 - Natural light
 - Complex community
 - Permeable sediments
 - Works with O_2 , temperature (other fast sensors)
 - In situ conditions
- 

Autonomous free falling deep sea landers

- Flotation unit
- Releasable ballast
- Chamber, Profiler or Eddy Covariation system
- Release ballast for positive buoyancy
 - Burn wire for programmed ascent
 - Acoustic release, redundant
 - Magnesium bolt for safety
- GPS transmitter
- Radio beacon
- Radar reflector



In situ lander and lab



- O_2 penetration underestimated in lab
- O_2 diffusive uptake overestimated in the lab

In situ data logging

Field Datalogger Mini

- 2 or 4 channels
- All Unisense sensors
- STOX support
- 300 or 6000 m depth rating
- Up to 20 days battery life
- Interface with CTD or other equipment
- 9.5 cm OD x 28 cm, 4.4 kg



Field Datalogger Mini – Data example

- H₂S and H₂ around Hydrothermal vents
- 2.4 to 3.3 km water depth
- Operated with ROV

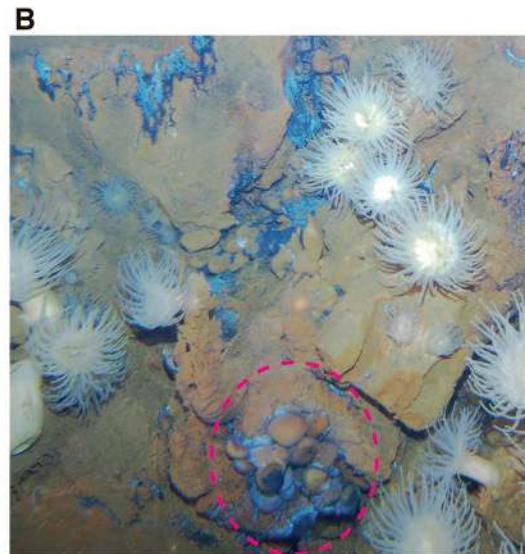
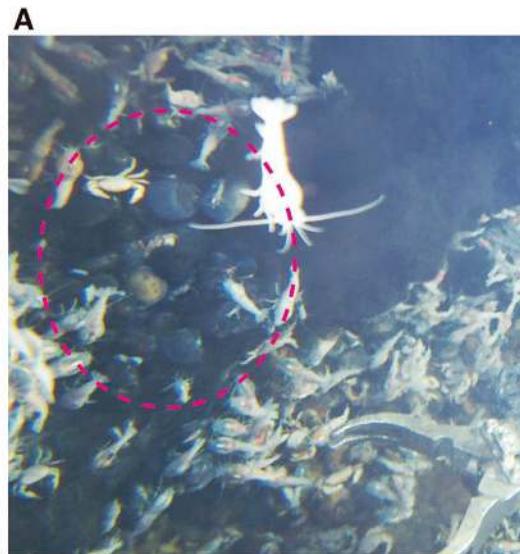


Table 1 Physical and chemical conditions of habitats of *Alviniconcha marisindica* populations (kAlv and eAlv populations) and *Chrysomallon squamiferum* in the Kairei and Edmond hydrothermal fields.

Sampling site	Temp. range [average] (°C)	DO range [average] (µM)	H ₂ range [average] (µM)	H ₂ S range [average] (µM)
Kairei field				
Reference bottom seawater	1.7–1.8 [1.8]	207	<0.03	<0.5
<i>A. marisindica</i> (kAlv) colony at the Monju chimney	11.9–60.0 [19.3]	104–166 [144]	20.1–44.6 [34.1] in situ	137–211 [185] in situ
			11.8–20.2 [16.5] onboard	<1.5 onboard
<i>C. squamiferum</i> colony at the Monju chimney	8.4–13.5 [12.6]	133–187 [158]	8.00–8.60 [8.40] in situ	23.5–29.6 [25.1] in situ
			6.37–10.2 [8.16] onboard	<1.5 onboard
Edmond field				
Reference bottom seawater	1.8–1.9 [1.9]	208	<0.03	<0.5
<i>A. marisindica</i> (eAlv) colony at the Giant shrimp chimney	6.5–38.1 [26.8]	147–202 [184]	0.39–0.45 [0.42] in situ	109–127 [120] in situ
			0.70–4.52 [2.52] onboard	19–26 [23.3] onboard

STOX sensor

- Switchable Trace Oxxygen sensor
- Trace amounts of O₂ can be measured reliably with a STOX sensor
- Detection limit down to 5-10 nM O₂



STOX sensor

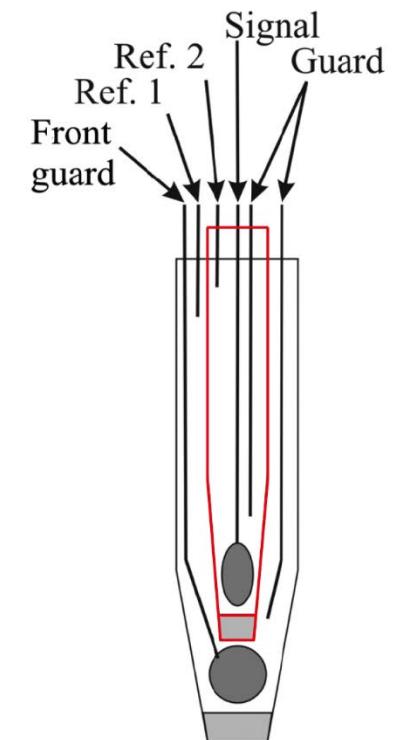
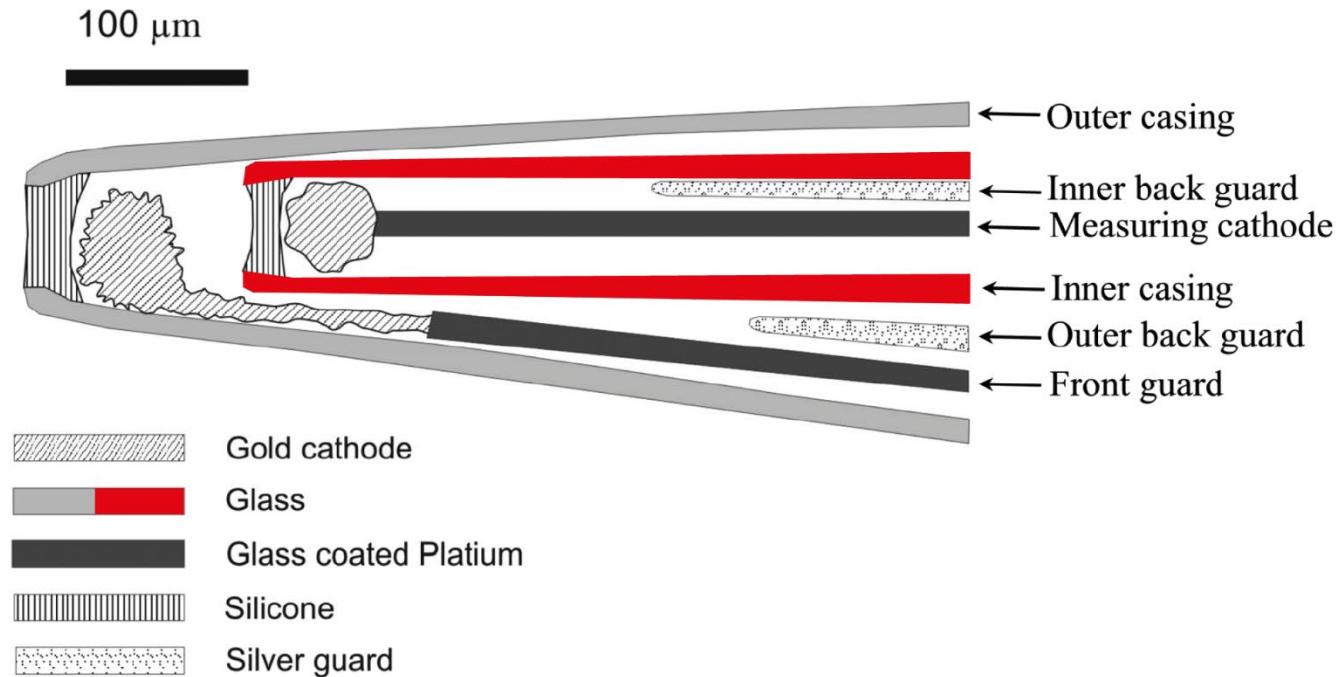
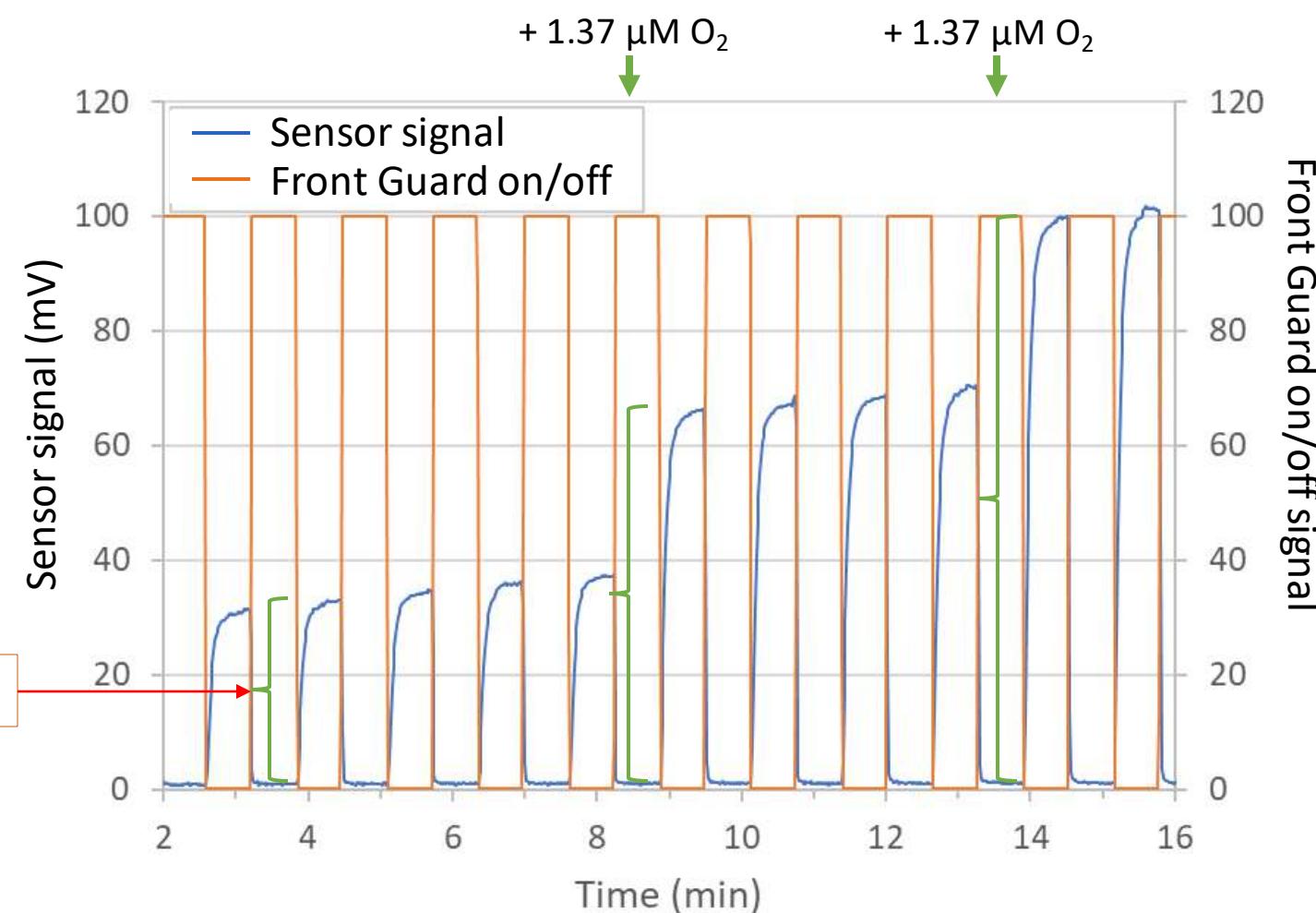


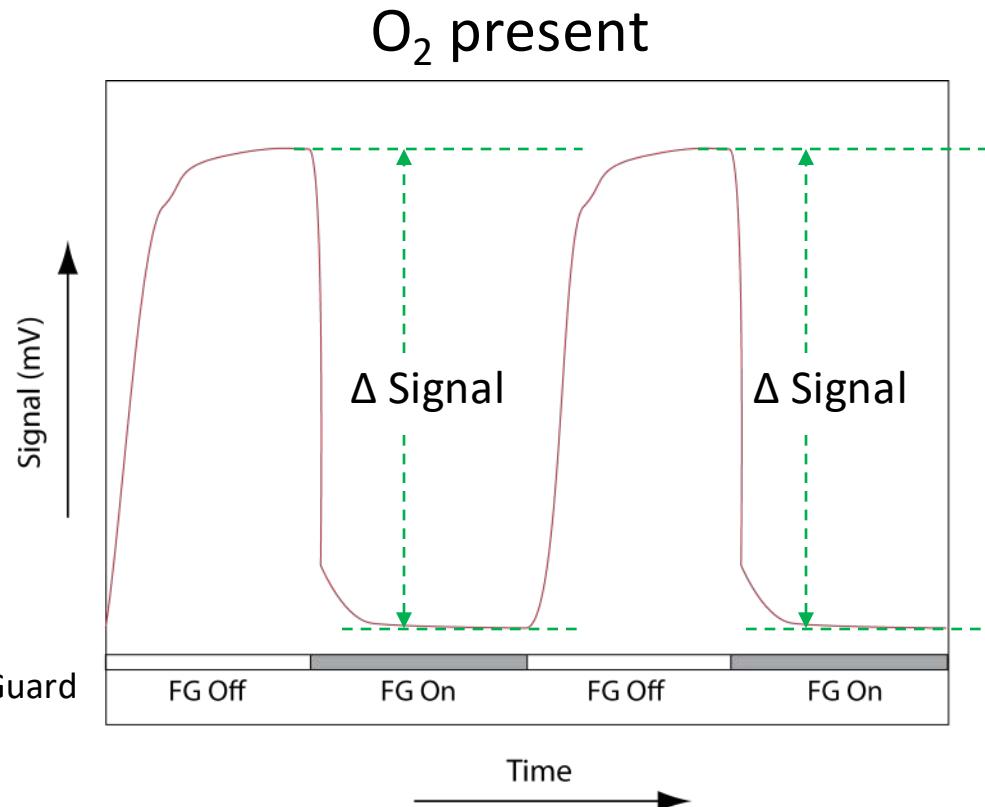
Fig. 1. Left: Schematic drawing of STOX microsensor tip. Not visible are the Ag/AgCl internal references in both inner and outer electrolyte reservoirs. Right: Diagram showing all anodes and cathodes in a STOX sensor.

STOX sensor - signal



$$\text{O}_2 \text{ signal} = \text{Sensor signal}_{\text{FG off}} - \text{Sensor signal}_{\text{FG on}} = \Delta \text{O}_2 \text{ signal}$$

STOX sensor



Δ Signal at zero O₂:

- Signal FG On = Signal FG Off
- Δ Signal = 0 mV

STOX sensor - calibration

At zero O_2 :

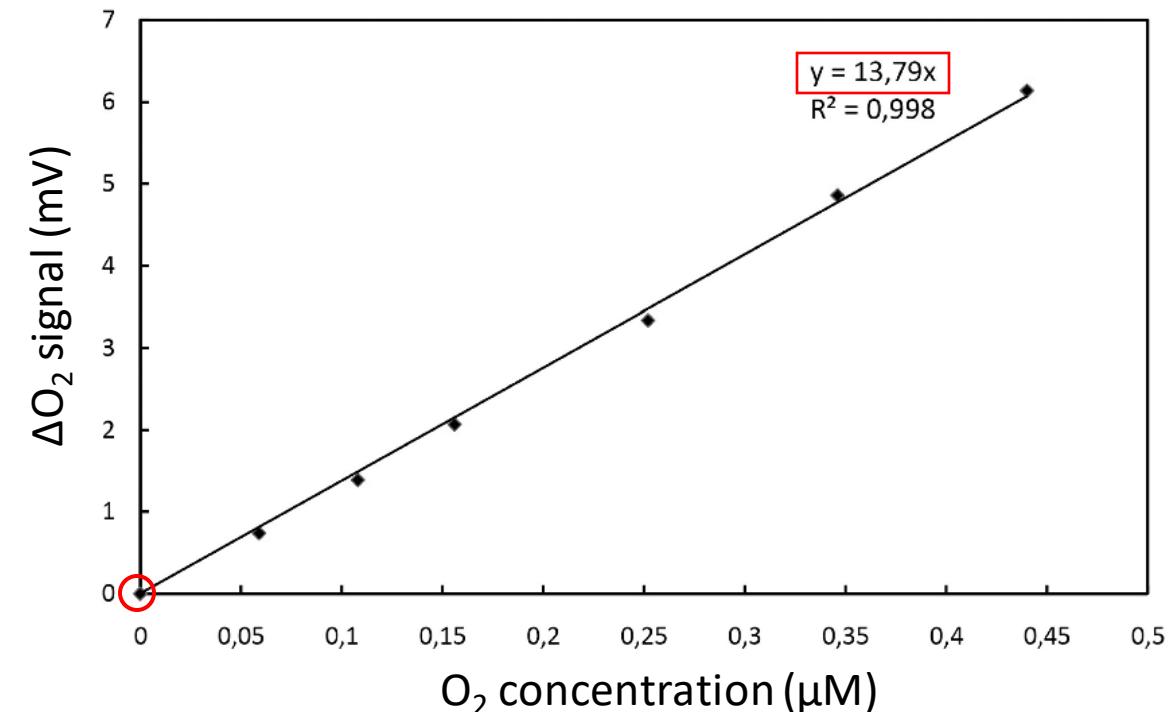
$$\text{Sensor signal}_{\text{FG off}} - \text{Sensor signal}_{\text{FG on}} = 0$$

Sensor response is linear

O_2 conc. is proportional to ΔO_2 signal

1-point calibration

$$O_2 \text{ conc.} = \Delta O_2 \text{ signal}/y$$



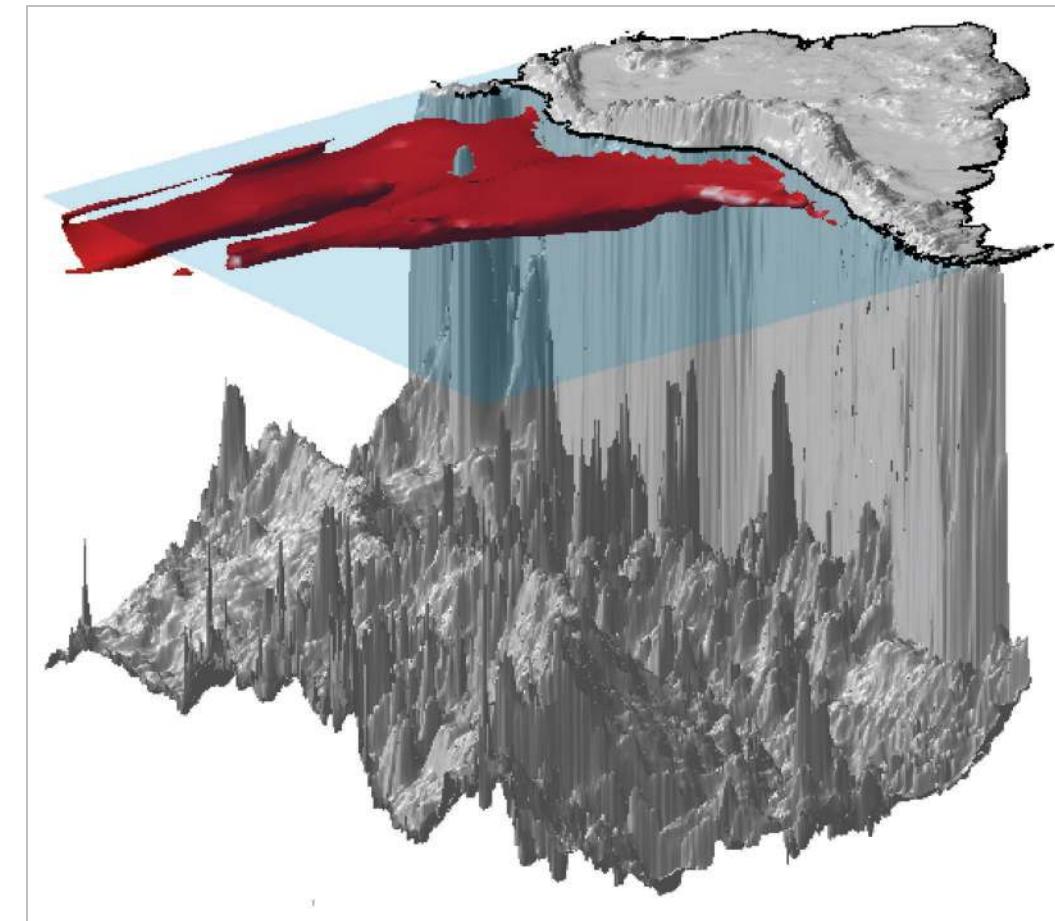
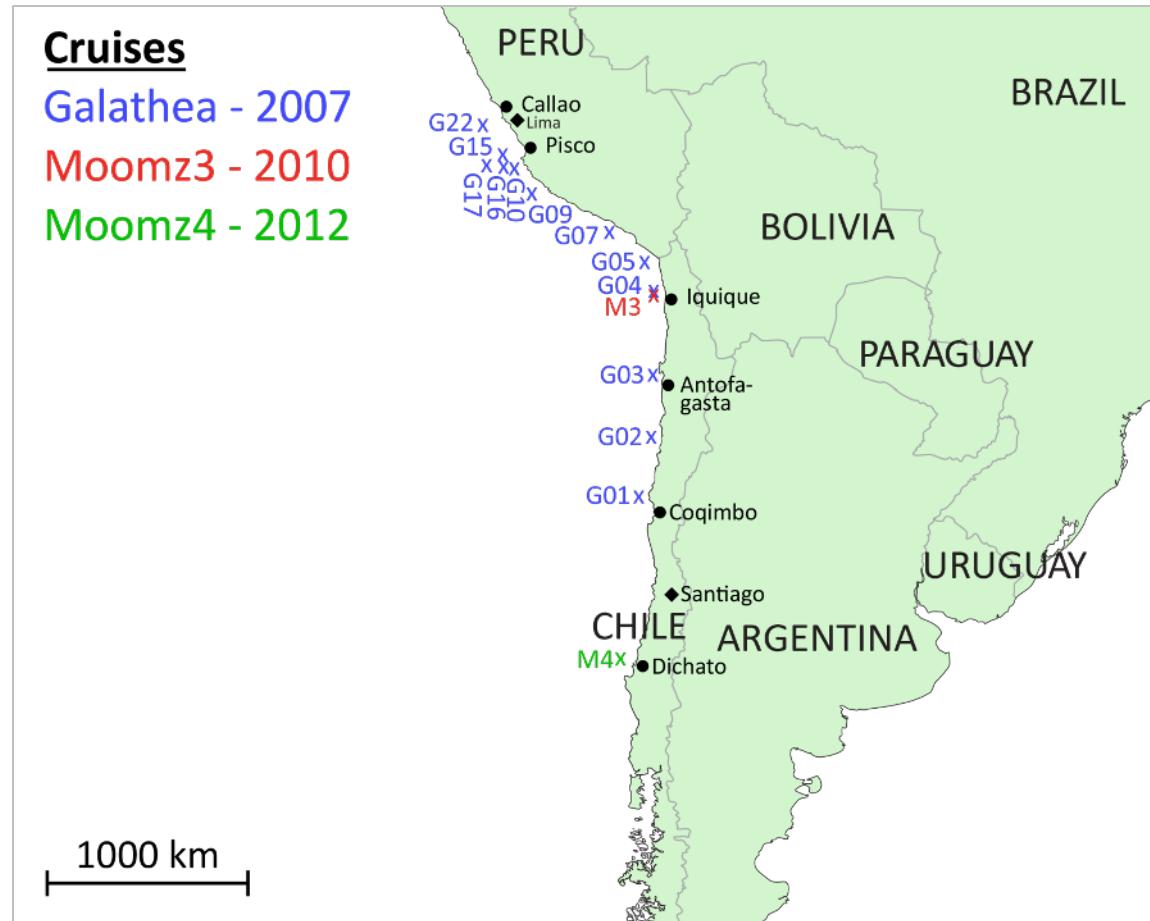
STOX sensor – Eastern South Pacific

Cruises

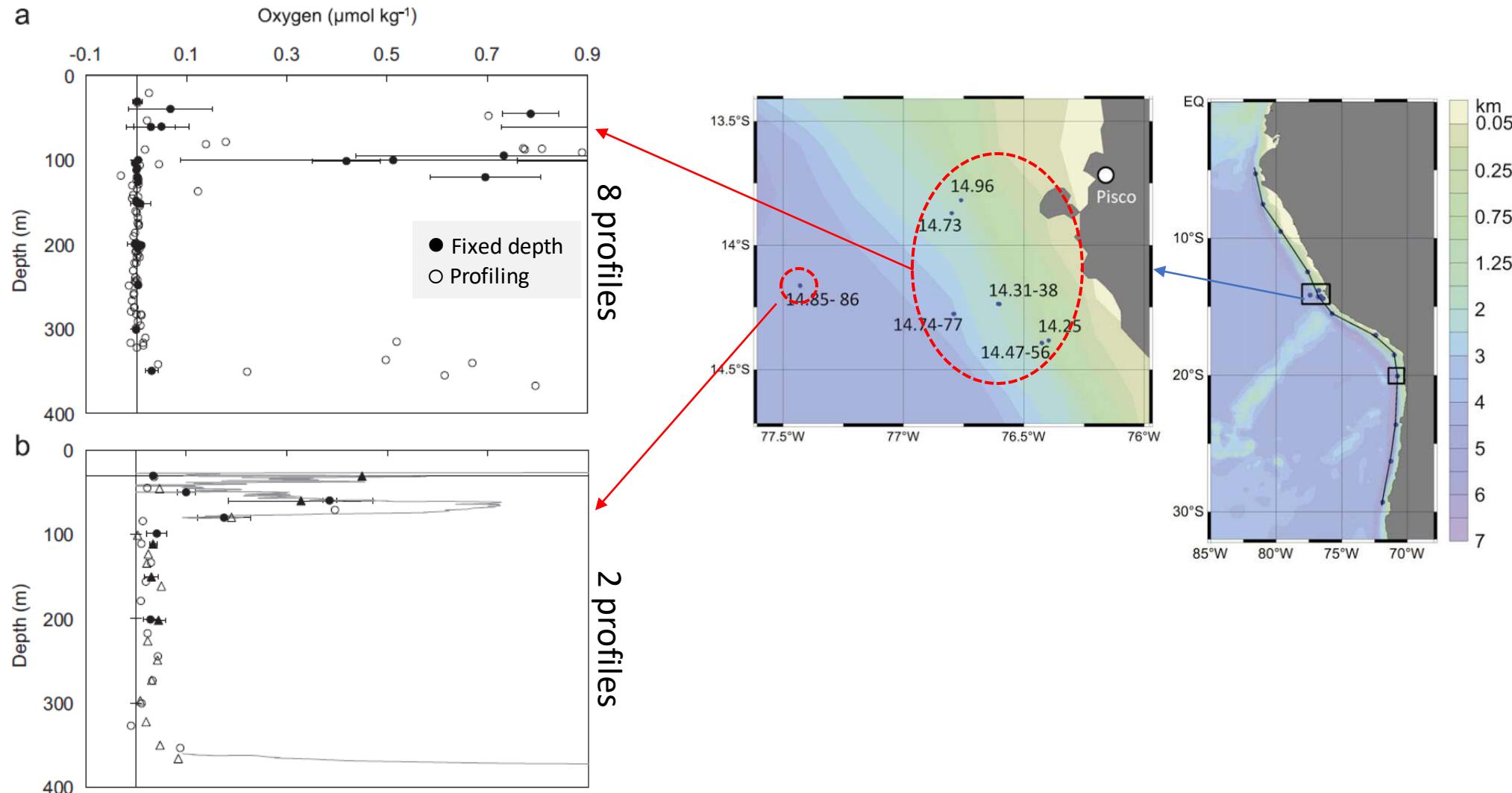
Galathea - 2007

Moomz3 - 2010

Moomz4 - 2012



STOX [O₂] – Along Chile and Peru

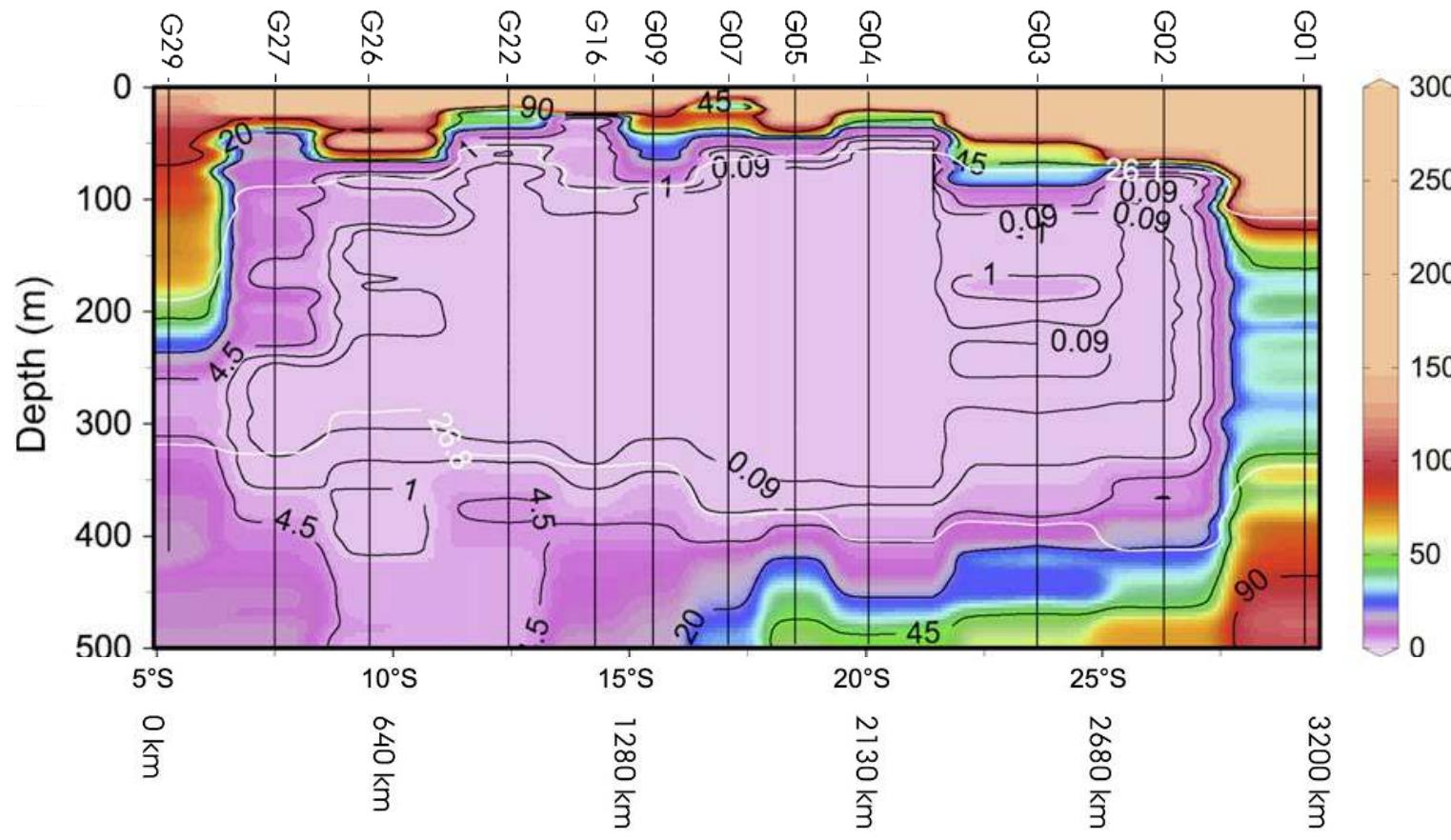


Thamdrup, B., T. Dalsgaard, and N. P. Revsbech. 2012. Widespread functional anoxia in the oxygen minimum zone of the eastern South Pacific. Deep Sea Res. Part I 65: 36–45.

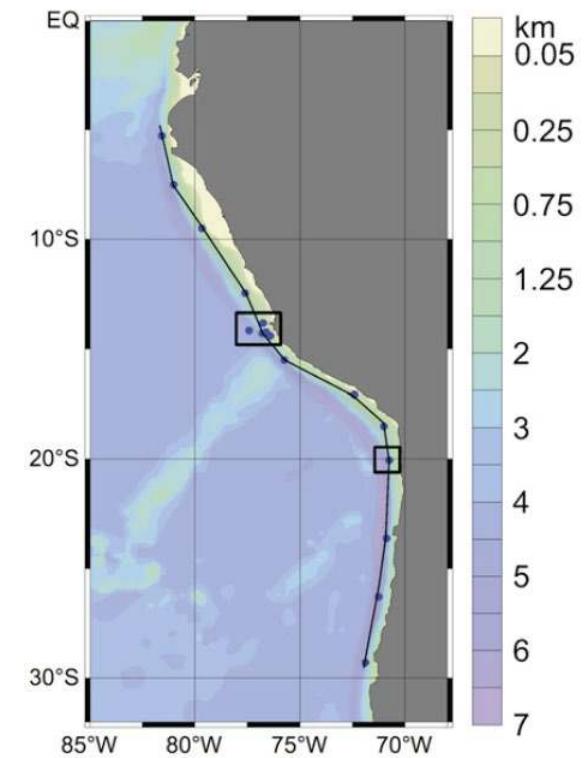


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STOX [O₂] – Along Chile and Peru



Detection limits:
STOX O₂ = 5 - 9 nM
SBE corrected = 90 nM

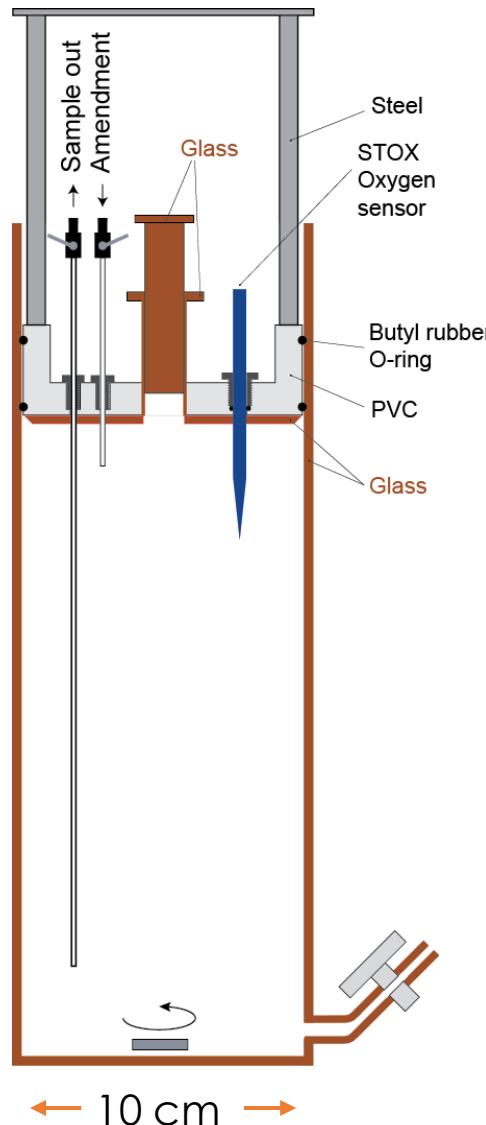


Thamdrup, B., T. Dalsgaard, and N. P. Revsbech. 2012. Widespread functional anoxia in the oxygen minimum zone of the eastern South Pacific. Deep Sea Res. Part I 65: 36–45.

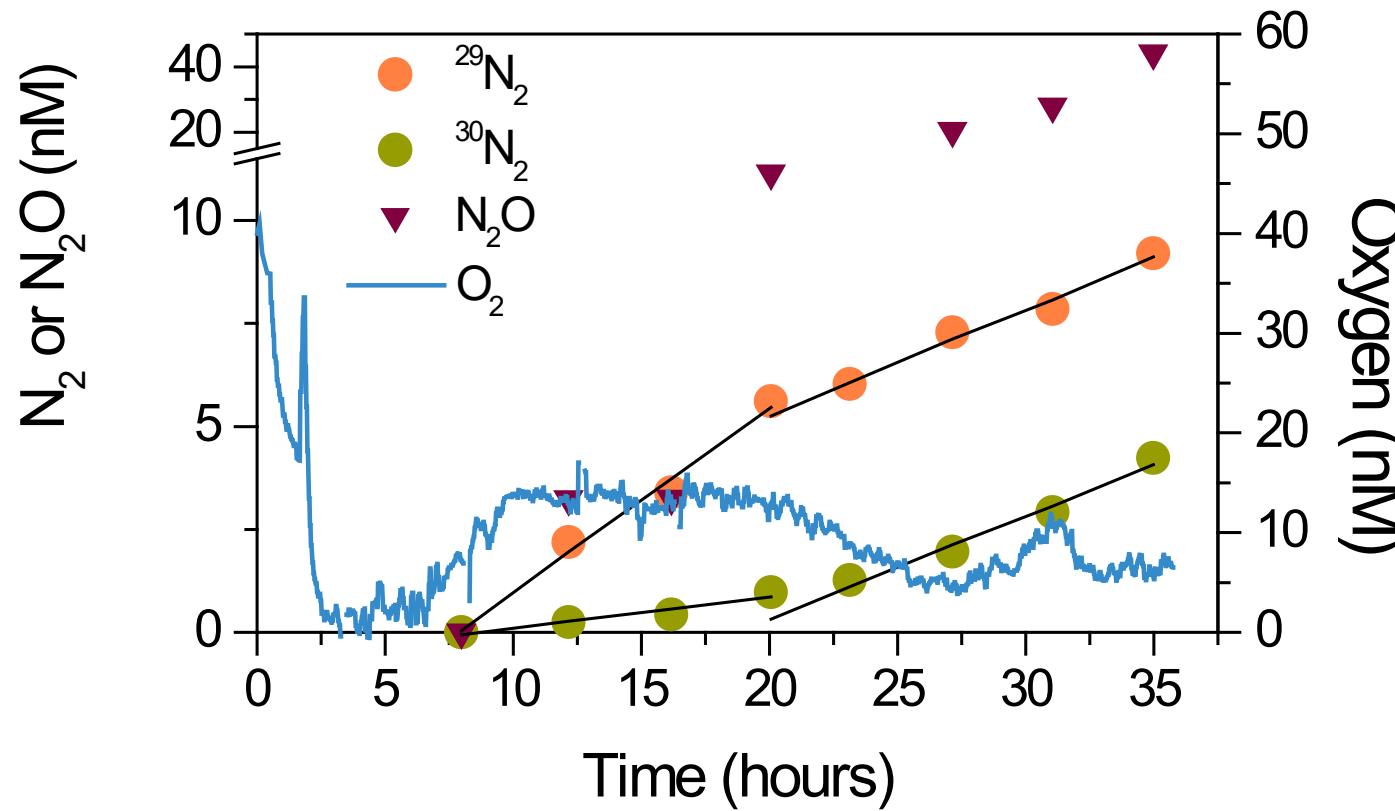
STOX sensor – Controlled reactor experiments

Reactor experiments

- Volume = 2 l
- ^{15}N tracer addition
 - Denitrification
 - Anammox
- Manipulate and monitor O_2



STOX sensor – Monitoring in reactor



$^{15}\text{NO}_2^-$ added:

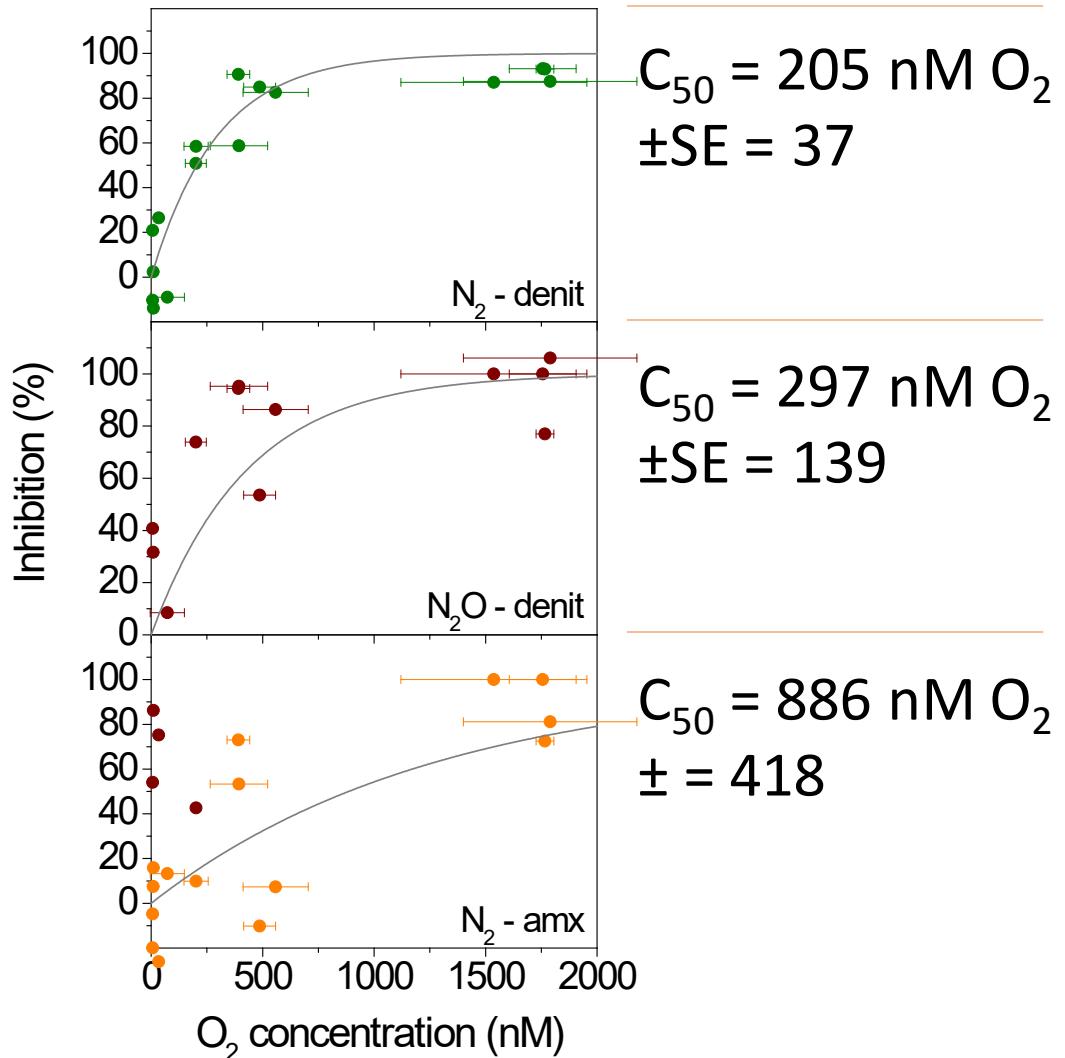
Anammox:



Denitrification:

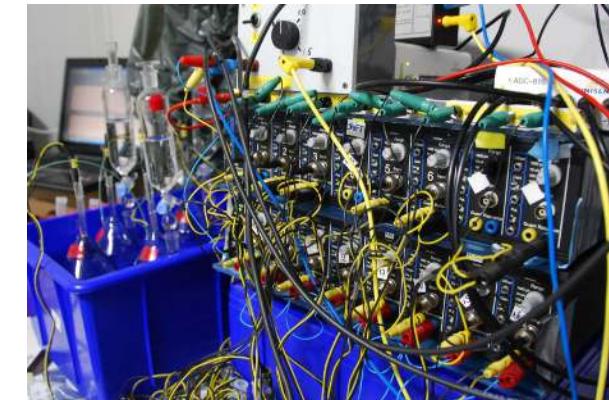


STOX - Inhibition by O₂



O₂ is important in the nM range

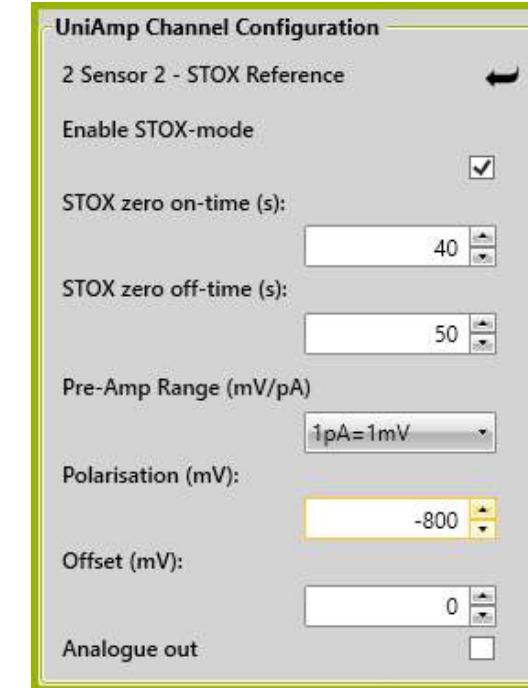
	Process	Product	C ₅₀ nM O ₂	K _m nM O ₂
Anaerobic	Denitrification	→ N ₂	205	
	Denitrification	→ N ₂ O	297	
	Anammox	→ N ₂	886	
Aerobic	NO ₃ ⁻ reduction	→ NO ₂ ⁻	740	
	NH ₄ ⁺ oxidation	→ NO ₂ ⁻		330
	NO ₂ ⁻ oxidation	→ NO ₃ ⁻		780



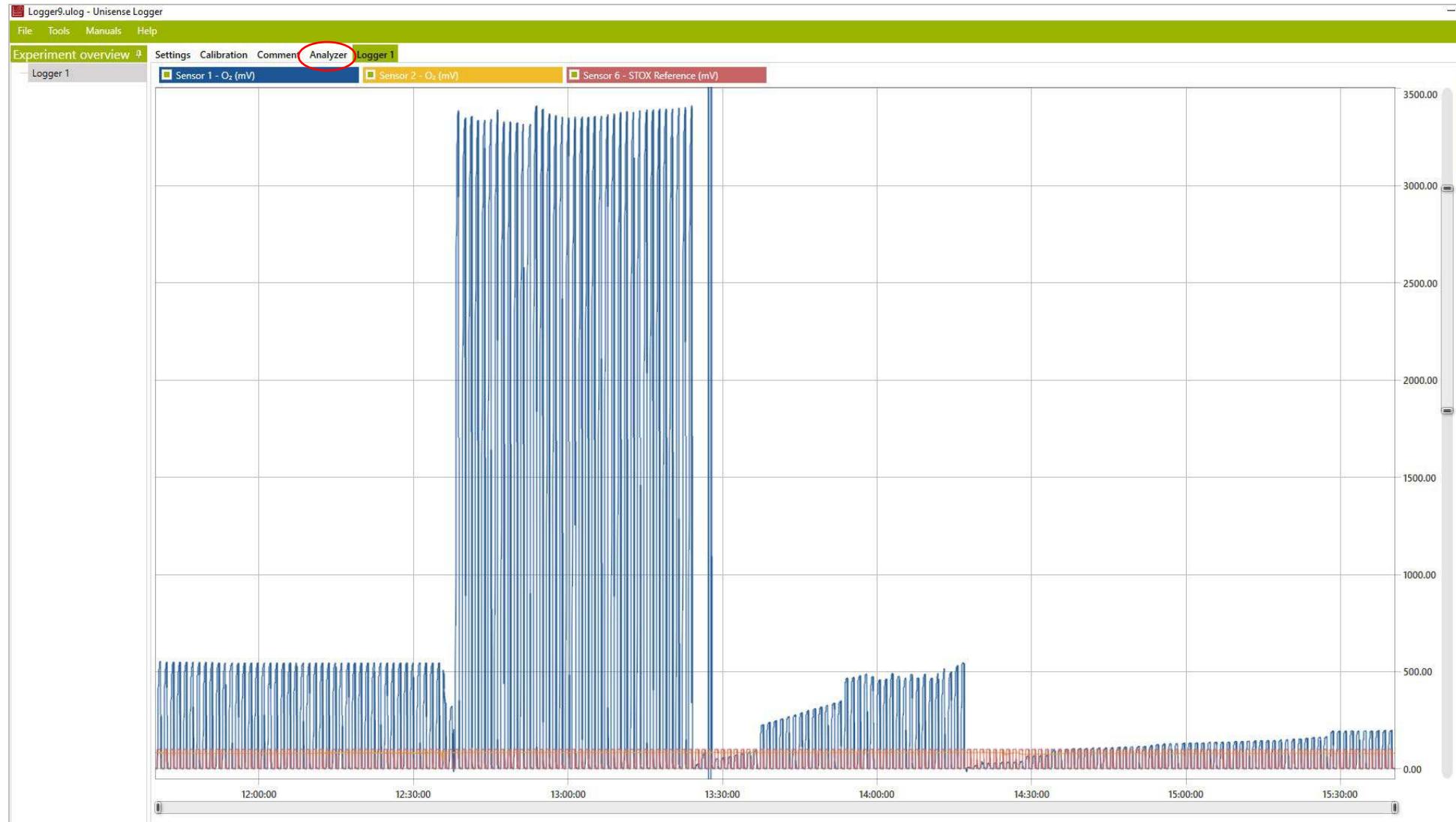
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Unisense STOX in the lab

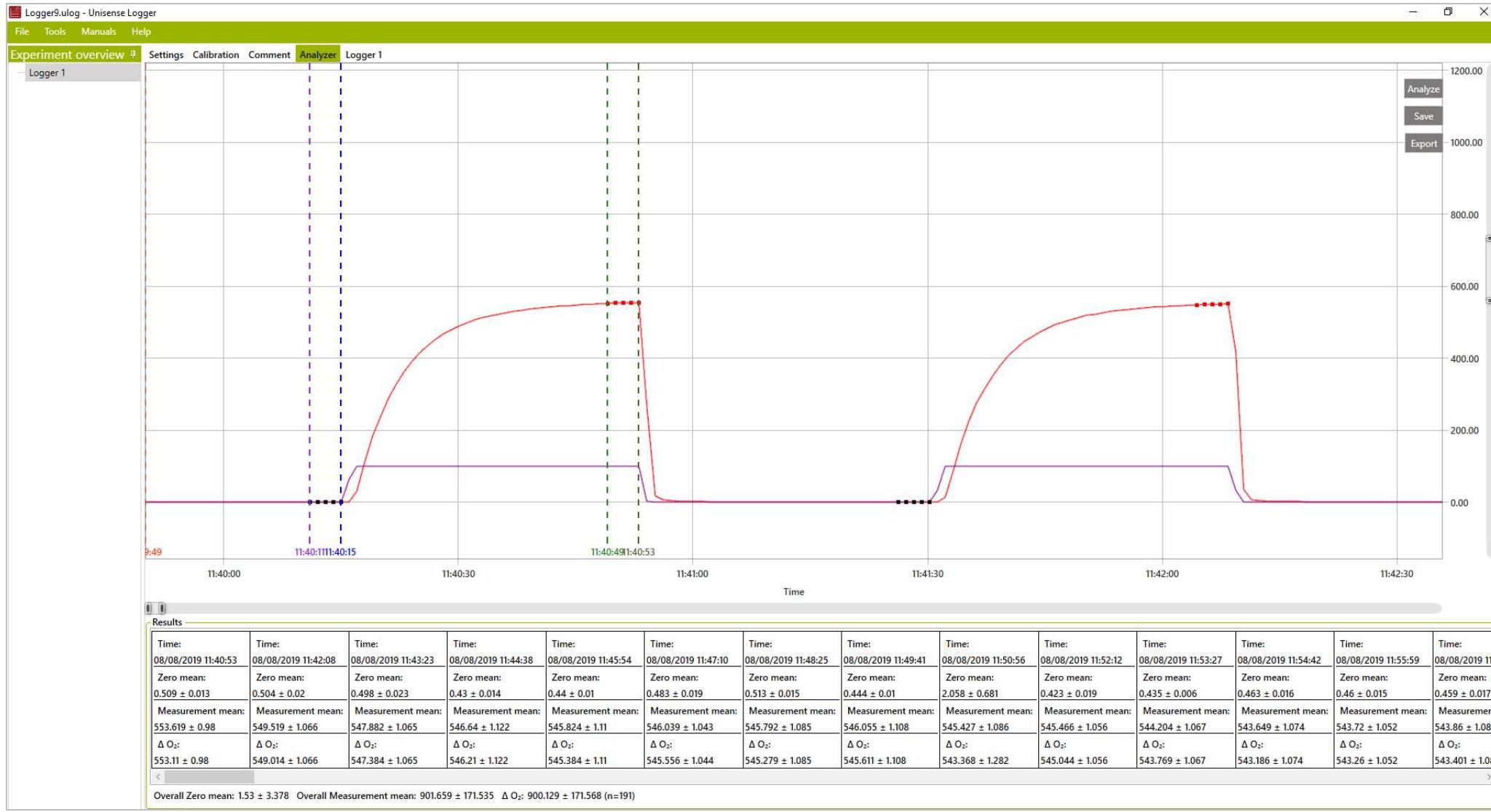
- UniAmp series of amplifiers has built in STOX support
- Plug and play
- STOX on/off timing adjustable in software



Unisense STOX data analysis



Unisense STOX data analysis



UNISENSE



A close-up photograph showing a clear plastic pipette tip positioned over a light-colored, granular sample, likely soil or a similar material. A small amount of liquid is being dispensed from the tip. The background is blurred, focusing attention on the pipette and the sample.

Time for questions !

Unisense Microsensor Academy:

<https://www.unisense.com/support/knowledge>

Contact us: sales@unisense.com